

# 1 DRIPS: Domain Randomisation for Image-based

## 2 Perivascular spaces Segmentation

3 Luna Bitar<sup>1,2</sup>, Mario Díaz<sup>3,4</sup>, Roberto Duarte Coello<sup>5,6</sup>, María d.C. Valdés-Hernández<sup>5,6</sup>,  
4 Hendrik Mattern<sup>1,7,8</sup>, Katja Neumann<sup>9</sup>, Malte Pfister<sup>9</sup>, Carolin Beck<sup>9</sup>, Huy Trong Mai<sup>9</sup>, Erelle  
5 Fuchs<sup>10</sup>, Serena Tang<sup>11,12</sup>, Duygu Tosun<sup>13</sup>, Bianca Besteher<sup>14,15,16</sup>, Tonia Rocktäschel<sup>14</sup>,  
6 Philipp A. Reuken<sup>17</sup>, Andreas Stallmach<sup>17</sup>, Nils Opel<sup>14,25</sup>, Christian Gaser<sup>14,15,16,18</sup>, Martin  
7 Walter<sup>14,15,16</sup>, Marc Dörner<sup>1,19</sup>, Philipp Arndt<sup>1,9</sup>, Daniel Behme<sup>10</sup>, Christiane Piechowiak<sup>20</sup>, Yves  
8 Lading<sup>20</sup>, Patrick Müller<sup>20</sup>, Rüdiger Braun-Dullaeus<sup>20</sup>, Sven G. Meuth<sup>21</sup>, for the Alzheimer's  
9 Disease Neuroimaging Initiative<sup>+</sup>, Joanna M. Wardlaw<sup>5,6</sup>, Stefanie Schreiber<sup>1,9</sup>, María Trujillo<sup>3</sup>,  
10 Emrah Düzel<sup>1,22</sup>, Gabriel Ziegler<sup>1,22</sup>, Jose Bernal<sup>1,5,22,23,\*</sup>

### 11 Author affiliations:

12 <sup>1</sup>German Centre for Neurodegenerative Diseases (DZNE), Magdeburg, Germany

13 <sup>2</sup>Institute of Medical Systems Bioinformatics (IMSB), Centre for Molecular Neurobiology (ZMNH), University  
14 Medical Centre Hamburg-Eppendorf (UKE), Hamburg, Germany

15 <sup>3</sup>Multimedia and Computer Vision (MCV) Lab, Universidad del Valle, Cali, Colombia

16 <sup>4</sup>Perception and Intelligent Systems (PSI) Research Group, School of Electrical and Electronics Engineering,  
17 Universidad del Valle, Cali, Colombia

18 <sup>5</sup>Institute for Neuroscience and Cardiovascular Research, Row Fogo Centre for Research into Ageing and The  
19 Brain, Department of Neuroimaging Sciences, The University of Edinburgh, Edinburgh, UK

20 <sup>6</sup>UK Dementia Research Institute Centre at the University of Edinburgh, Edinburgh, UK

21 <sup>7</sup>Center for Behavioural Brain Sciences (CBBS), Magdeburg, Germany

22 <sup>8</sup>Biomedical Magnetic Resonance, Otto-von-Guericke University, Magdeburg, Germany

23 <sup>9</sup>Department of Neurology, University Hospital Magdeburg, Magdeburg, Germany

24 <sup>10</sup>Clinic for Neuroradiology, University of Magdeburg, Magdeburg, Germany

25 <sup>11</sup>Department of Bioengineering, University of California San Francisco, San Francisco, CA, USA

26 <sup>12</sup>Department of Bioengineering, University of California Berkeley, Berkeley, CA, USA

27 <sup>13</sup>Department of Radiology and Biomedical Imaging, University of California San Francisco, San Francisco, CA,  
28 USA

29 <sup>14</sup>Centre for Intervention and Research on adaptive and maladaptive brain Circuits underlying mental health (C-I-  
30 R-C), Halle-Jena-Magdeburg

31 <sup>15</sup>Department of Psychiatry and Psychotherapy, Jena University Hospital, Jena, Germany

32 <sup>16</sup>German Centre for Mental Health (DZPG), partner site Halle-Jena-Magdeburg, Germany

33 <sup>17</sup>Department of Internal Medicine IV, Gastroenterology, Hepatology and Infectious diseases, Jena University  
34 Hospital

35 <sup>18</sup>Department of Neurology, Jena University Hospital, Jena, Germany

36 <sup>19</sup>Department of Consultation-Liaison-Psychiatry and Psychosomatic Medicine, University Hospital Zurich, Zurich,  
37 Switzerland

38 <sup>20</sup>Division of Cardiology and Angiology, University Hospital Magdeburg, Magdeburg, Germany

39 <sup>21</sup>Department of Neurology, Heinrich-Heine-University, Düsseldorf, Germany

40 <sup>22</sup>Institute of Cognitive Neurology and Dementia Research, Otto-von-Guericke University Magdeburg, Magdeburg,  
41 Germany

42 <sup>23</sup>Cerebrovascular Imaging and Research Lab, Department Artificial Intelligence in Biomedical Engineering (AIBE),  
43 Friedrich-Alexander-Universität Erlangen-Nürnberg (FAU), Erlangen, Germany

44 <sup>24</sup>Department of Psychiatry & Neuroscience, Campus Benjamin Franklin, Charité Universitätsmedizin Berlin,  
45 Germany

46 <sup>25</sup>German Centre for Mental Health (DZPG), partner site Berlin-Potsdam, Germany

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52 \*Corresponding author

53

54 Correspondence to:

55 Jose Bernal

NOTE: This preprint reports new research that has not been certified by peer review and should not be used to guide clinical practice.

56 Full address: Cerebrovascular Research and Imaging Lab, Department of Artificial Intelligence in  
57 Biomedical Engineering (AIBE), Friedrich-Alexander Universität Erlangen-Nürnberg (FAU),  
58 Nürnberg Strasse 74, 91052 Erlangen, Germany  
59 E-Mail: [jose.bernal@fau.de](mailto:jose.bernal@fau.de)

## 60 Abbreviations

61	AUPRC	Area under the precision-recall curve
62	BG ROI	Basal ganglia region of interest
63	CSF	Cerebrospinal fluid
64	CSO ROI	Centrum semiovale region of interest
65	DSC	Dice similarity coefficient
66	DRIPS	Domain Randomisation for Image-based PVS Segmentation
67	FFT	Fast Fourier transformation
68	IFFT	Inverse Fast Fourier transformation
69	MRI	Magnetic resonance imaging
70	PVS	Perivascular spaces
71	ROC	Receiver operating characteristic curve
72	ROI	Region of interest
73	RORPO	Ranking the orientation responses of path operators
74	SNR	Signal-to-noise ratio
75	SVF	Stationary velocity field
76	TE	Echo time
77	TR	Repetition time
78	WMH	White matter hyperintensities

## 79 Abstract

80 Perivascular spaces (PVS) are emerging as sensitive imaging markers of brain health.  
81 Yet, accurate out-of-sample PVS segmentation remains challenging since existing  
82 methods are modality-specific, require dataset-specific tuning, or rely on manual labels  
83 for (re-)training. We propose DRIPS (Domain Randomisation for Image-based PVS  
84 Segmentation), a physics-inspired framework that integrates anatomical and shape  
85 priors with a physics-based image generation process to produce synthetic brain  
86 images and labels for on-the-fly deep learning model training. By introducing variability  
87 through resampling, geometric and intensity transformations, and simulated artefacts,  
88 it generalises well to real-world data. We evaluated DRIPS on MRI data from five  
89 cohorts spanning diverse health conditions ( $N = 165$ ; T1w and T2w, isotropic and  
90 anisotropic imaging) and on a 3D ex vivo brain model reconstructed from histology.  
91 We evaluated its performance using the area under the precision–recall curve  
92 (AUPRC) and Dice similarity coefficient (DSC) against manual segmentations and  
93 compared it with classical and deep learning methods, including Frangi, RORPO,  
94 SHIVA-PVS, and nnU-Net. Only DRIPS and Frangi achieved AUPRC values above  
95 chance across all cohorts and the ex vivo model. On isotropic data, DRIPS and nnU-  
96 Net performed comparably, outperforming the next-best method by a median of  
97  $+0.17\text{--}0.39$  AUPRC and  $+0.09\text{--}0.26$  DSC. On anisotropic data, DRIPS outperformed  
98 all competitors by a median of  $+0.13\text{--}0.22$  AUPRC and  $+0.07\text{--}0.14$  DSC. Importantly,  
99 its performance was not associated with white matter hyperintensity burden. DRIPS  
100 delivers accurate, fully automated PVS segmentation across heterogeneous imaging  
101 settings, reducing the need for manual labels, modality-specific models, or cohort-  
102 dependent tuning.

103 **Keywords:** Perivascular spaces; Segmentation; Domain Randomisation; Deep  
104 Learning; Magnetic Resonance Imaging

## 105 1 Introduction

106 Perivascular spaces (PVS) are anatomical passageways that surround arterioles,  
107 capillaries, and venules in the brain and an integral part of the neurovascular unit  
108 (Gouveia-Freitas and Bastos-Leite, 2021; Wardlaw et al., 2020). Collectively, PVS  
109 form a brain-wide network of conduits for cerebrospinal fluid (CSF) circulation  
110 (Hirschler et al., 2025; Wardlaw et al., 2020, 2009; Yamamoto et al., 2024), a function  
111 that underlies the clearance of metabolic and neurotoxic waste products (Braun and  
112 Iliff, 2020; Hablitz and Nedergaard, 2021; Iliff et al., 2014, 2012; Mestre et al., 2018;  
113 Rasmussen et al., 2018; Wardlaw et al., 2020). These spaces are dynamic, with the  
114 capacity to shrink and enlarge, at times reaching a calibre that renders them visible *in*  
115 *vivo* on magnetic resonance imaging (MRI) at standard clinical field strengths (1.5 T /  
116 3 T) (Kern et al., 2023; Kim et al., 2023; Lynch et al., 2023; Menze et al., 2024; Vikner  
117 et al., 2022). PVS enlargement is pathological (Bown et al., 2022; Francis et al., 2019;  
118 Okar et al., 2023; Wardlaw et al., 2020) and is considered an early structural change  
119 of impaired cerebrovascular and brain waste clearance function (Francis et al., 2019;  
120 Ineichen et al., 2022; Okar et al., 2023; Schreiber et al., 2023; Wardlaw et al., 2020;  
121 Waymont et al., 2024).

122 The growing recognition of PVS as a non-invasive imaging marker of compromised  
123 brain health function has prompted the development and large-scale deployment of  
124 computational methods for their quantification and monitoring (Smith et al., 2019;  
125 Waymont et al., 2024). Broadly, the literature describes two strategies: classical and  
126 machine learning based methods (Waymont et al., 2024). Classical methods use the  
127 morphology and CSF-like signal of PVS to distinguish them from other brain structures  
128 and, when multimodal data are available, from other concomitant lesions, such as

129 white matter hyperintensities (WMH) and lacunar infarcts (Ballerini et al., 2020, 2018;  
130 Barisano et al., 2025; Barnes et al., 2022; Bernal et al., 2021b, 2020; Boespflug et al.,  
131 2018; Duarte Coello et al., 2024; Menze et al., 2024; Schwartz et al., 2019; Valdés  
132 Hernández et al., 2024). These well-established methods offer high sensitivity (Bernal  
133 et al., 2022)—a double-edged sword that often necessitates careful parameter tuning  
134 and post-processing to minimise false positives (Ballerini et al., 2018; Bernal et al.,  
135 2022, 2020; Valdés Hernández et al., 2024). Machine learning methods, on the other  
136 hand, leverage supervised learning (Boutinaud et al., 2021a; Cai et al., 2024; Chai et  
137 al., 2025; Dubost et al., 2019a, 2019b; González-Castro et al., 2016; Hou et al., 2017;  
138 Lian et al., 2018; Park et al., 2016; Pham et al., 2024; Rashid et al., 2023; Zhang et  
139 al., 2017). Within this category, deep learning has emerged as the most widely  
140 adopted method (Waymont et al., 2024). The main advantage of deep learning is that,  
141 with sufficiently large, diverse, and well-annotated datasets, models are able to  
142 overcome some of the limitations of classical strategies. Nonetheless, the scarcity of  
143 such datasets (Sudre et al., 2024) generally hinders their ability to generalise  
144 effectively to unseen datasets (Billot et al., 2023a; Chalcroft et al., 2025). This, in turn,  
145 constrains their broader applicability beyond their training sets.

146 Domain randomisation has emerged as an alternative to address this generalisation  
147 problem (Tobin et al., 2017). In contrast to data augmentation—which applies  
148 predefined spatial and intensity transformations to existing images—domain  
149 randomisation uses procedural image generation models, conditioned on  
150 segmentations with fully randomised parameters, to create synthetic datasets for  
151 training deep learning models. The diversity of training samples enables models  
152 trained with domain randomisation to learn domain-independent features that  
153 characterise target structures well. SynthSeg is an example of a successful method

154 taking advantage of domain randomisation (Billot et al., 2023a). It is a model that  
155 segments brain structures on real MRI acquired with diverse sequences and  
156 modalities without retraining, despite being trained exclusively on synthetic data. Since  
157 its introduction in the early 2020s, approaches leveraging domain randomisation have  
158 been successfully applied to a variety of tasks, including skull-stripping (Hoopes et al.,  
159 2022), segmentation of brain structures (Billot et al., 2023a, 2023b), WMH (Laso et  
160 al., 2023), and stroke lesions (Chalcroft et al., 2025), as well as super-resolution  
161 (Iglesias et al., 2023) and image registration (Hoffmann et al., 2024).

162 Realism in synthetic data generation is not essential; rather, it is crucial that generated  
163 data pose challenges comparable to real-world scenarios, enabling networks to learn  
164 robust and transferable features (Billot et al., 2023a). For synthetic PVS data  
165 generation, Bernal et al. (2022b) developed an open-source physics-inspired  
166 computational model that creates 3D digital reference objects containing PVS-like  
167 structures distributed throughout the brain. The generation process involved inserting  
168 randomly oriented tubular structures into a high-resolution head model, followed by k-  
169 space sampling, motion artefact simulation, and Rician noise corruption to produce  
170 low-resolution T2w-like images. Although it was originally conceived for method  
171 benchmarking, this computational model may serve as a basis for data generation  
172 and, when combined with domain randomisation, may facilitate training of deep  
173 learning algorithms with improved generalisability (Bernal et al., 2022).

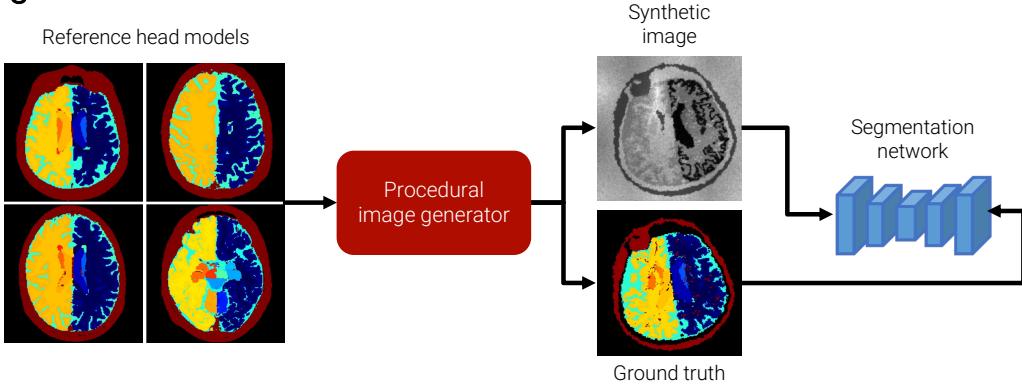
174 Here, we introduce DRIPS (Domain Randomisation for Image-based PVS  
175 Segmentation), the first physics-inspired domain randomisation framework specifically  
176 developed for accurate out-of-sample PVS segmentation. DRIPS accurately  
177 segmented PVS in imaging data acquired with multiple imaging sequences and  
178 resolutions from patients with varying health conditions. It performed robustly across

179 all these settings and frequently surpassed both classical image-processing and deep  
180 learning methods.

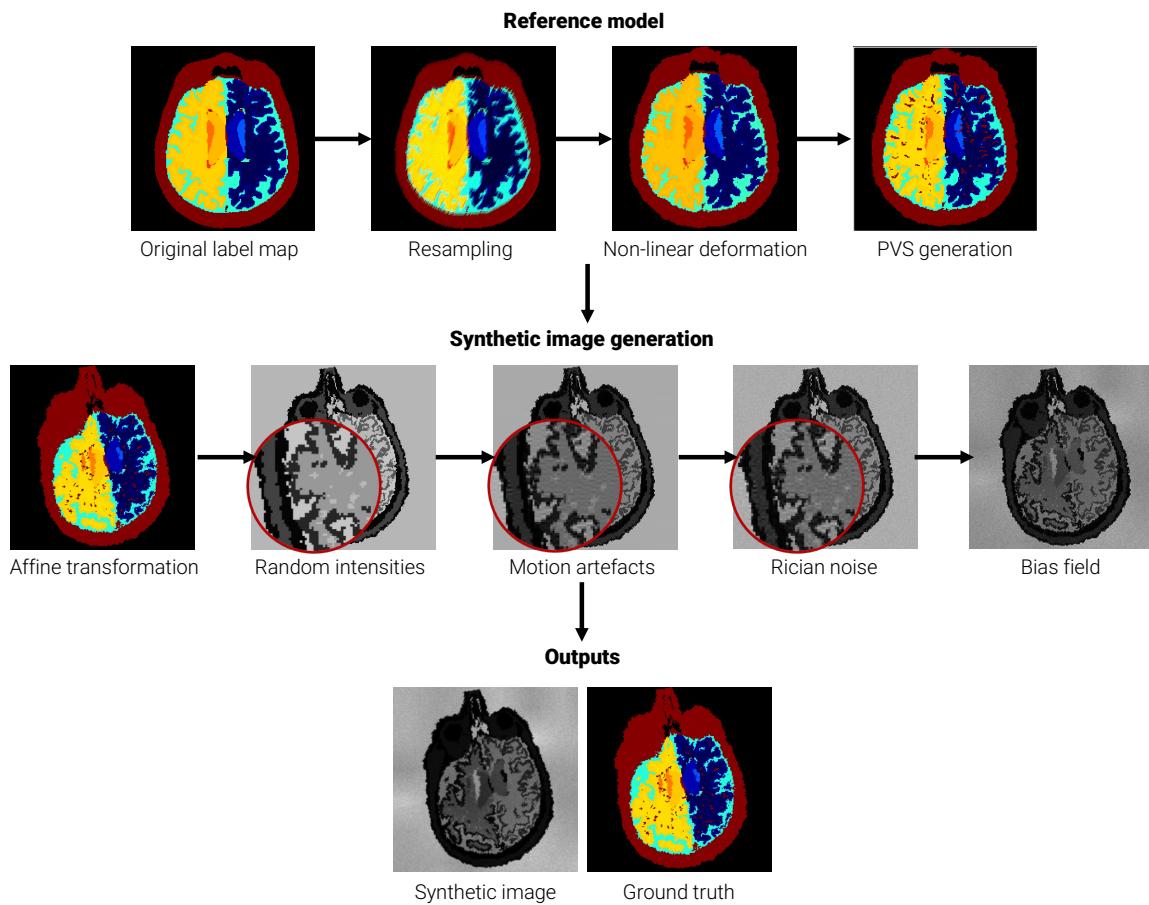
181 **2 DRIPS**

182 DRIPS is a domain randomisation framework specifically designed for out-of-sample  
183 PVS segmentation (Figure 1). It integrates anatomical and shape priors of the human  
184 head and PVS with a physics-inspired procedural image generation process to create  
185 synthetic brain images and corresponding label maps. It then uses these synthetic  
186 datasets, generated on the fly, to train segmentation networks. By introducing  
187 variability through random resampling, geometric transformations, intensity sampling,  
188 and simulated MR artefacts, DRIPS produces models that achieve high segmentation  
189 accuracy and generalise effectively to real-world data. The following sections provide  
190 detailed descriptions of each step.

### (A) DRIPS



### (B) Procedural image generator



191

192 **Figure 1. Schematic of DRIPS.** (A) DRIPS is a domain randomisation framework that trains  
193 segmentation networks for out-of-sample PVS segmentation. It combines anatomical and shape priors  
194 of the human head and PVS with physics-inspired image generation to create synthetic brain images  
195 and corresponding label maps containing PVS-like structures. It then trains segmentation networks—  
196 here exemplified with a U-Net—on these synthetic image–label pairs. By exposing networks to broad  
197 imaging variability during training, DRIPS achieves accurate PVS segmentation across diverse cohorts,  
198 modalities, and acquisition settings. (B) Starting from anatomical head atlases with added synthetic,  
199 tortuous PVS-like structures, DRIPS procedurally generates heterogeneous synthetic brain images  
200 through random resampling, non-linear and affine transformations, intensity sampling, and typical MR  
201 image corruptions and artefacts (motion artefacts, Rician noise, bias fields). Red-circled regions in the  
202 procedural image generator correspond to zoomed-in views.

## 203 **2.1 Reference model**

### 204 **2.1.1 Head model**

205 We used 840 three-dimensional atlases derived from T1w and FLAIR scans of the  
206 ADNI database and CSVD Magdeburg cohorts as head models. Each atlas was a  
207 segmentation map with  $1 \text{ mm}^3$  resolution, in which every voxel was assigned to a  
208 specific class, including the lateral ventricles, white matter, WMH, cortical grey matter,  
209 cerebral white matter, cerebellar grey matter, brain stem, subcortical structures, or  
210 extracranial structures (Billot et al., 2023a). We used SynthSeg (Billot et al., 2023a)  
211 and LST-AI (Wiltgen et al., 2024) to obtain whole-brain parcellations and WMH masks,  
212 respectively. To introduce further anatomical variability, we applied random nonlinear  
213 diffeomorphic deformations to the original set of atlases. Specifically, we sampled a  
214 small stationary velocity field (SVF;  $10 \times 10 \times 10 \times 3$ ) from a zero-mean Gaussian  
215 distribution, with standard deviation  $\sigma_{SVF}$  randomly drawn from a uniform distribution.  
216 The range of  $\sigma_{SVF}$  was set from 0 to 4 to allow for varying degrees of deformation. We  
217 then upsampled this field to full image resolution using trilinear interpolation to obtain  
218 a high-resolution SVF. Finally, we warped the original label map with this deformation  
219 field using nearest-neighbour interpolation to produce deformed brain atlases.

### 220 **2.1.2 PVS model**

221 We then added synthetic PVS-like structures to the generated head models. Although  
222 PVS are commonly described as tubular in clinical studies (Wardlaw et al., 2020), they  
223 do not conform to strictly Euclidean shapes and often exhibit tortuous geometries  
224 (Bernal et al., 2022). To capture this non-Euclidean morphology and have flexibility in

225 representing PVS-like structures, we modelled them as tortuous tubular structures  
226 using the following parametric equation:

227 
$$x(t) = 0, y(t) = \cos(\alpha t), z(t) = t,$$

228 where  $t \sim \mathcal{U}(t_{low}, t_{high})$  and  $\alpha \sim \mathcal{U}(\alpha_{low}, \alpha_{high})$  control the length and tortuosity of  
229 the generated PVS. Longer and more tortuous PVS structures are obtained by  
230 increasing  $t$  and decreasing  $\alpha$ . We allowed  $t$  to vary between 2 and 10 voxels and  $\alpha$   
231 between 1/10 to 1/5. We placed these synthetic PVS in random locations within the  
232 white matter (normal-appearing and hyperintensities) and subcortical grey matter  
233 regions. We aligned each PVS towards the lateral ventricles, and to prevent clustering  
234 near the brain's centre, we used a stratified jittered sampling strategy.

235 **2.2 Procedural synthetic image generation**

236 We developed a procedural image generation model to create synthetic images for  
237 training the segmentation network. Using the head and PVS models, we generated  
238 synthetic images on the fly with fully randomised parameters, varying image  
239 intensities, contrasts, resolutions, and artefacts within each batch. The individual steps  
240 for synthetic data generation are illustrated in Figure 1 and described in detail below:

241 **2.2.1 Resampling and voxel size variability**

242 To enable the model to process scans acquired at different voxel sizes, we generated  
243 synthetic images and label maps with varying voxel sizes. We achieved this by  
244 resampling the input label maps to a randomly selected target voxel size. The target  
245 voxel size was randomly chosen on-the-fly during training, with each dimension  
246 varying between 0.5 mm and 4 mm to enable the processing of both research and

247 clinical scans. We resampled label maps using nearest-neighbour interpolation to  
248 preserve the original discrete voxel values.

249 **2.2.2 Affine transformations**

250 We applied random affine transformations to the label maps to increase anatomical  
251 variability and, at the same time, to preserve structural integrity. Rotation, scaling,  
252 shearing, and translation parameters were randomly selected, with all values sampled  
253 from uniform distributions (see (Billot et al., 2023a) for more information).

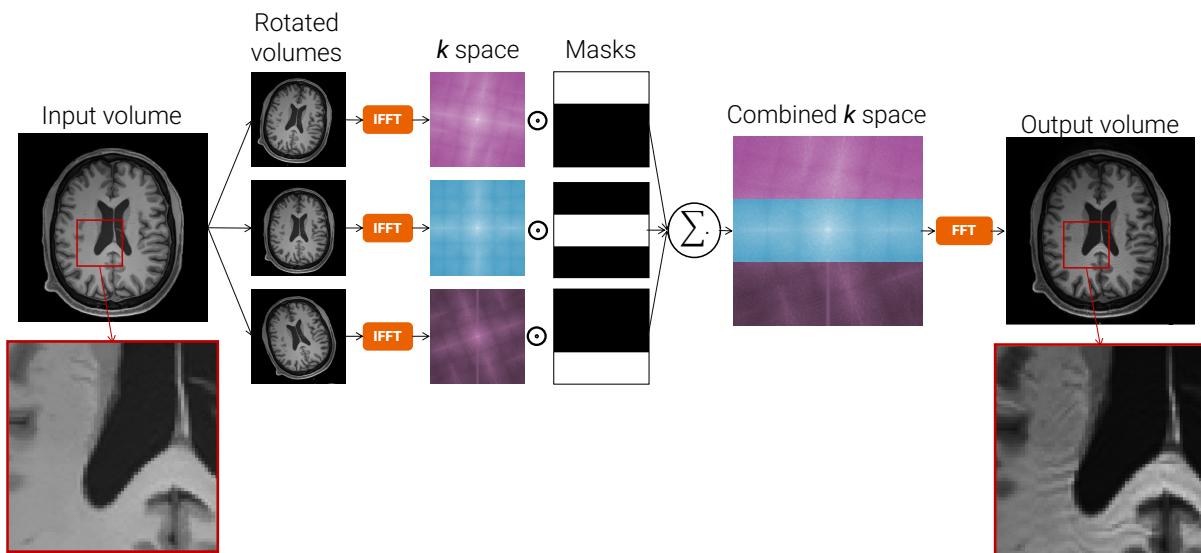
254 **2.2.3 Random intensity generation**

255 We assigned each anatomical structure a single random intensity, sampled from a  
256 standard uniform distribution  $\mathcal{U}(0, 1)$ . This procedure varied structure intensities  
257 across images, eliminating consistent local patterns and forcing the model to rely on  
258 shape and spatial information for segmentation.

259 **2.2.4 Motion artifacts**

260 Motion artefacts are a common source of image degradation in MRI and can markedly  
261 affect the visibility and quantification of fine structures such as PVS (Bernal et al.,  
262 2022). Owing to their thin, elongated morphology, PVS are particularly susceptible to  
263 being mistaken for motion streaks, making artefact mitigation especially critical for this  
264 application. We simulated rotational motion during  $k$ -space acquisition using a  
265 composite  $k$ -space model (Bernal et al., 2022, 2021a; Shaw et al., 2020).  
266 We first rotated the original synthetic volume twice by random angles within  $[-15^\circ, 15^\circ]$   
267 around random axes and compute the  $k$ -space of both the original and rotated  
268 volumes. We then generated a composite  $k$ -space by taking between 50% and 100%  
269 of the data from the original volume and replacing the remainder with data from the

270 rotated volumes along a randomly selected axis. Finally, we transformed the resulting  
271 composite *k*-space to image space to produce a motion-corrupted image. The level of  
272 displacement between consecutive frames and the time at which the motion occurs  
273 determines the severity and appearance of the motion artefacts in the resulting image  
274 (Figure 2).



275

276 **Figure 2. Simulation of motion artefacts in DRIPS.** Motion was modelled in *k*-space by combining  
277 data from the original and randomly rotated volumes along a chosen axis. Varying the fraction of *k*-  
278 space segments taken from the original and rotated versions and the "timing" of motion yielded different  
279 levels of blurring and ghosting. Abbreviation: IFFT/FFT: (inverse) fast Fourier transformation

## 280 2.2.5 Rician noise

281 MRI data are inherently affected by noise originating during acquisition in *k*-space,  
282 where additive white Gaussian noise affects both the real and imaginary components  
283 of the complex signal (Gudbjartsson and Patz, 1995). Following transformation into  
284 the spatial domain and magnitude reconstruction, this noise takes a Rician distribution.  
285 To add Rician-distribution noise to the images, we added uncorrelated additive white  
286 Gaussian noise to the real and imaginary channels of the combined *k* space. The  
287 Gaussian noise standard deviation was computed as  $\sigma_{noise} = \mu_{signal}/10^{SNR_{dB}/20}$ , with

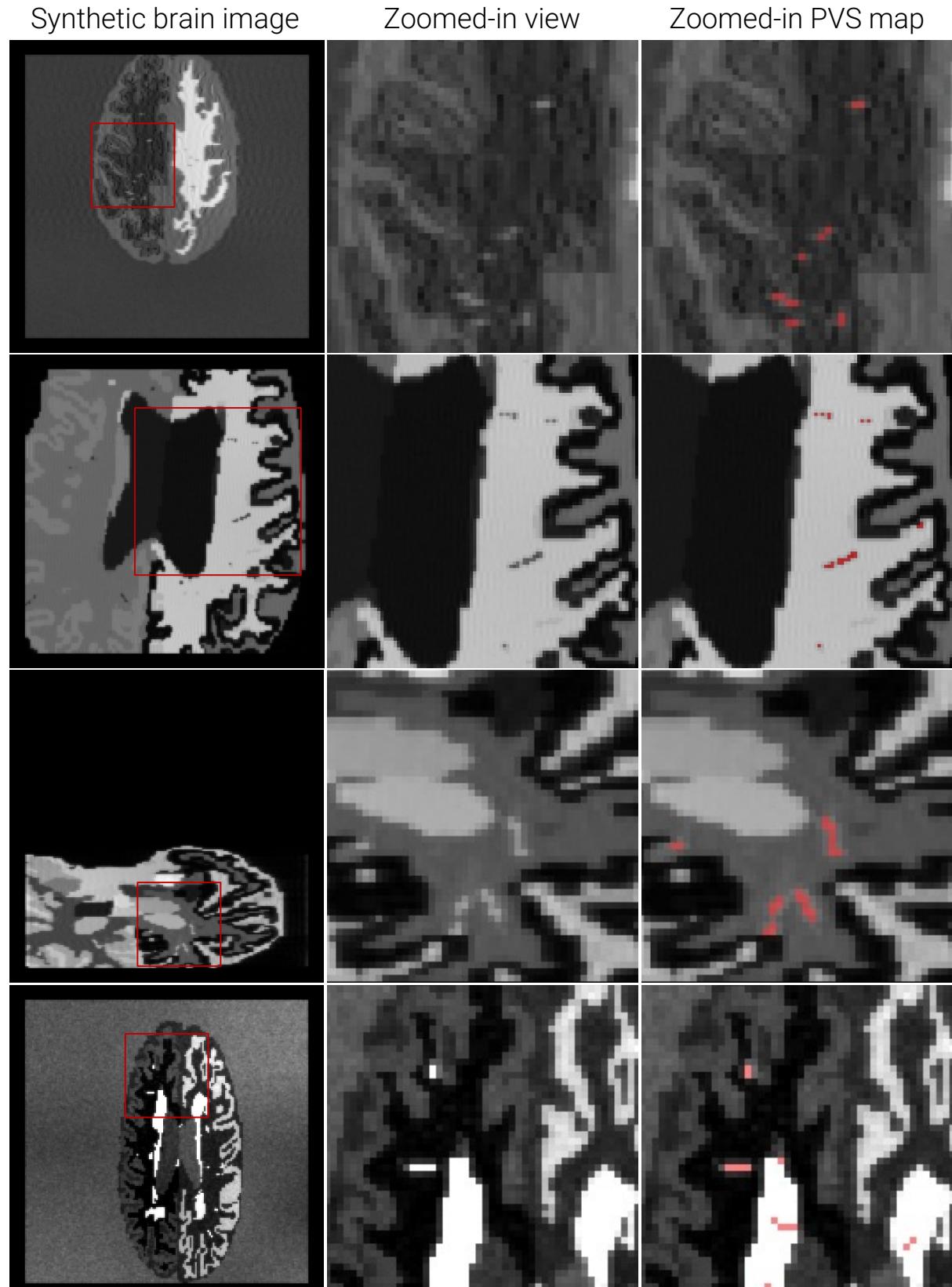
288 the SNR in decibels sampled from a uniform distribution  $\mathcal{U}(SNR_{min}, SNR_{max})$ . We set  
289  $SNR_{min}$  and  $SNR_{max}$  to 5 dB and 40 dB to simulate a broad spectrum of image noise.

290 **2.2.6 Bias field inhomogeneity**

291 We modelled bias field corruption to mimic MRI intensity inhomogeneities arising from  
292 B-field inhomogeneities and magnetic field variations. Following the approach in (Billot  
293 et al., 2023a), we sampled a  $4 \times 4 \times 4$  Gaussian random volume  $\sigma_B$ , upsampled it to  
294 image resolution for smooth variation, exponentiated to enforce positive multiplicative  
295 effects, and applied it to the synthetic image. We normalised intensities to [0, 1] and  
296 subjected the image to a random Gamma transformation to introduce additional non-  
297 linear signal variations.

298 **2.2.7 Final training pair**

299 Figure 3 presents four examples of synthetically generated images with their  
300 corresponding label maps, illustrating the variability in brain shape, structure, and  
301 intensity introduced by DRIPS. These DRIPS-generated pairs can be used as input  
302 and ground truth for training segmentation models.



303

304 **Figure 3. Synthetic brain images with corresponding ground truths, obtained using the**  
305 **proposed domain randomisation method.** Note that synthetic images vary, among other aspects, in  
306 anatomy, orientation, intensity, PVS distribution, and levels of Rician noise, motion artefacts, and  
307 inhomogeneities.

## 308 2.3 DRIPS-based model training and testing

309 DRIPS provides a basis for training segmentation networks for out-of-sample PVS  
310 segmentation. In this work, we employed a 3D U-Net as the segmentation model, a  
311 well-established architecture capable of capturing both local and global spatial  
312 features critical for accurate segmentation. We note that the framework is architecture-  
313 agnostic and can be readily adapted to alternative architectures.

314 The 3D U-Net consisted of five encoding and five decoding levels. Each level had two  
315 convolutional layers with kernels of size  $3 \times 3 \times 3$ , followed by a batch normalisation  
316 layer and max pooling or upsampling layers, depending on whether the level was part  
317 of the encoding or decoding part, respectively. All convolutional layers employed an  
318 exponential linear unit activation, except for the final layer, which had a softmax  
319 activation. The number of kernels per level doubled after each max pooling and halved  
320 after each upsampling layer. The first layer contained 24 feature maps. The network  
321 had skip connections to transfer feature maps from the encoding path to the decoding  
322 path.

### 323 2.3.1 Training on synthetic data

324 We trained the segmentation network in DRIPS for 50 epochs, each comprising 5000  
325 batches of size 1, with each image–label pair generated on the fly by the procedural  
326 image generator. We used the Adam optimiser (learning rate of  $10^{-4}$ ) and a generalised  
327 Dice loss function for model optimisation. The generalised Dice loss function for  
328 multiple classes is given by (Milletari et al., 2016):

$$329 \text{Generalised Dice Loss}(\text{GT}, S) = 1 - \sum_{c \in \{0,1\}} \frac{2 \cdot \sum_{x,y,z} \text{GT}_c(x, y, z) S_c(x, y, z)}{\sum_{x,y,z} \text{GT}_c(x, y, z)^2 + S_c(x, y, z)^2},$$

330 where  $c \in \{0,1\}$  denotes the considered classes (0: background, 1: PVS), and  $GT_c$  and  
331  $S_c$  are the ground truth and soft probability map for class  $c$ , respectively. We  
332 implemented the segmentation model in Keras with a TensorFlow backend. Training  
333 took approximately twelve days on an NVIDIA A100 Tensor Core GPU.

334 **2.3.2 Testing on real data**

335 To evaluate the model on real data, we first normalised image intensities before  
336 feeding the scans into the network (Billot et al., 2023a). Inference took approximately  
337 ten seconds on an NVIDIA A100 Tensor Core GPU and 60 seconds on CPU.

338 Contrast agnosticism encourages models to prioritise shape over intensity. Though  
339 advantageous for generalisation, this technique makes models prone to detecting  
340 “tubular” structures regardless of whether their intensity profiles match those of PVS.  
341 For example, although PVS appear hypointense in T1w imaging, sections of the  
342 internal and external capsules—which are not hypointense in this modality—were  
343 sometimes flagged as potential PVS (non-zero response). To restrict detection to  
344 hypointense structures in T1w images and hyperintense structures in T2w images, we  
345 thus applied the Laplacian operator during post-processing, retaining regions with  
346 positive and negative Laplacian values, respectively.

347 **3 Evaluation on real data**

348 **3.1 Cohorts and ground truth**

349 We tested DRIPS on images and manual PVS segmentations from 165 participants  
350 from five cohorts: post-COVID Brain (PCB; N=42) (Besteher et al., 2022), EBBIVD  
351 (N=18), heart failure with preserved ejection fraction on cerebral microangiopathy

352 (HIM; N=39; DRKS00031583) (Müller et al., 2024), MagDeburger DrAinage-Reserve-  
353 Score (MD-DARS; N=6) (Neumann et al., 2022), and ADNI-3 (N=60). Further  
354 information can be found in Table 1. Ethical approval was granted by the Ethics  
355 Committees of the University Hospital Magdeburg for the EBBIVD, HIM, and MD-  
356 DARS cohorts, and by the Ethics Committee of Jena University Medical School for the  
357 PCB cohort, and by Institutional Review Boards of all participating centres for the  
358 ADNI-3 cohort. All participants provided written informed consent in accordance with  
359 the Declaration of Helsinki.

360 Data used in the preparation of this article were obtained from the Alzheimer's Disease  
361 Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). The ADNI was launched  
362 in 2003 as a public-private partnership. The original goal of ADNI was to test whether  
363 serial magnetic resonance imaging, positron emission tomography, other biological  
364 markers, and clinical and neuropsychological assessment can be combined to  
365 measure the progression of mild cognitive impairment and early Alzheimer's disease.  
366 The current goals include validating biomarkers for clinical trials, improving the  
367 generalizability of ADNI data by increasing diversity in the participant cohort, and to  
368 provide data concerning the diagnosis and progression of Alzheimer's disease to the  
369 scientific community. For up-to-date information, see adni.loni.usc.edu

370 **Table 1. Imaging protocols, and PVS and WMH burden across cohorts.** The table summarises the  
371 imaging sequences, acquisition parameters, and scanner specifications used for manual PVS  
372 segmentation across the PCB, EBBIVD, HIM, MD-DARS, and ADNI cohorts. We also report PVS and  
373 WMH burden separately for the BG and CSO ROIs, presented as median counts, volumes, and  
374 fractional volumes, with interquartile ranges in brackets. Fractional volumes represent the volume of  
375 PVS within a region of interest relative to the volume of the region.

	<b>PCB</b>	<b>EBBIVD</b>	<b>HIM</b>	<b>MD-DARS</b>	<b>ADNI</b>
<b>N</b>	42	18	39	6	60
<b>Clinical groups</b>	Normal cognition post-COVID syndrome	Hypertensive arteriopathy Cerebral amyloid angiopathy	Individuals with heart failure with preserved ejection fraction	Individuals spanning the Alzheimer's disease continuum	Normal cognition Mild cognitive impairment Alzheimer's disease
<b>Imaging</b>					

Sequence for PVS segmentation	3D T1w MPRAGE	2D T2w TSE	2D T2w TSE	2D T2w TSE	3D T1w MPRAGE
Key parameters	TR = 2400 ms TE = 2.22 ms FA = 9°	TE = 73 ms TR = 6500 ms	TE = 73 ms TR = 6500 ms	TE = 73 ms TR = 6500 ms	TE = min full TR = 2300 ms TI = 900 ms FA = 9°
Voxel size [mm <sup>3</sup> ]	0.8 × 0.8 × 0.8	0.5 × 0.5 × 2.0	0.5 × 0.5 × 2.0	0.5 × 0.5 × 2.0	1.0 × 1.0 × 1.0
Magnetic field strength [T]	3	3	3	3	3
Scanner model/vendor	Siemens Tim Trio (Siemens Healthineers, Erlangen, Germany)	Skyra (Siemens Healthineers, Erlangen, Germany)	Skyra (Siemens Healthineers, Erlangen, Germany)	Skyra (Siemens Healthineers, Erlangen, Germany)	Siemens, GE, and Philips (multi-vendor)
<b>PVS burden</b>					
BG PVS count	397 [311–506]	890 [643–1368]	656 [499–1053]	816 [526, 1033]	36 [17–67]
BG PVS volume [ml]	0.263 [0.212–0.342]	0.779 [0.592–1.255]	0.587 [0.435–0.928]	0.699 [0.459, 0.992]	0.051 [0.022–0.099]
Fractional BG PVS volumes (%)	0.587 [0.416–0.611]	1.865 [1.394–2.768]	1.172 [0.945–1.839]	1.585 [1.118–2.046]	0.117 [0.052–0.232]
CSO PVS count	1370 [782–2450]	7692 [5204–9056]	6379 [5012–8227]	6424 [3943, 7143]	510 [267, 858]
CSO PVS volume [ml]	0.976 [0.545–1.686]	8.471 [5.854–9.552]	6.853 [5.266–8.691]	6.9705 [4.275, 7.397]	0.900 [0.479, 1.4305]
Fractional CSO PVS volumes (%)	0.344 [0.192–0.652]	3.027 [1.852–3.801]	2.200 [1.678–2.822]	2.446 [1.468–2.579]	0.387 [0.216–0.598]
<b>WMH burden</b>					
BG WMH volume [ml]	No FLAIR imaging	No FLAIR imaging	0.218 [0.017–0.837]	0.293 [0.032–1.537]	0.408 [0.103–0.878]
CSO WMH volume [ml]	No FLAIR imaging	No FLAIR imaging	1.510 [0.243–7.573]	2.081 [0.155–9.819]	3.364 [1.374–11.382]

376

377 Under the guidance of experienced neuroradiologists, four medical residents and one  
 378 neuroscientist segmented PVS manually using either Mango or ITK-SNAP. PVS  
 379 segmentation was performed on T1w scans for PCB and ADNI, and on T2w scans for  
 380 EBBIVD, HIM, and MD-DARS, following STRIVE criteria (Duering et al., 2023). The  
 381 smallest available paint tool was used to manually delineate PVS across all axial slices  
 382 throughout the entire brain. FLAIR sequences were taken into account, when  
 383 available, to minimise the inclusion of WMH.

384 **3.2 Evaluation metrics**

385 We assessed PVS segmentation using voxel-wise and lesion-wise Dice similarity  
 386 coefficients ( $DSC_{voxel}$  and  $DSC_{lesion}$ ) and the area under the precision–recall curve  
 387 (AUPRC).  $DSC_{voxel}$  quantifies spatial overlap between the predicted and ground-truth

388 binary maps within the ROI.  $DSC_{lesion}$  evaluates object-wise agreement after  
389 connected-component labelling, measuring overlap between individual predicted and  
390 reference PVS (e.g., one-inside-the-another criterion) (Maier-Hein et al., 2024).  
391 AUPRC summarises segmentation performance across all possible thresholds. We  
392 opted for precision–recall over receiver operating curves given the pronounced class  
393 imbalance (Maier-Hein et al., 2024).

394 Since all of our evaluations are performed out-of-sample, discrepancies may arise  
395 between how PVS were segmented in the training data and how they appear in an  
396 unseen dataset (e.g. where PVS boundaries end). To mitigate this potential mismatch  
397 and ensure a fair comparison across methods, we derived DSC values by thresholding  
398 each output at the operating point on the precision–recall curve that maximised  
399 segmentation performance. In practice, this corresponds to the threshold at which the  
400 trade-off between sensitivity and precision yields the highest  $DSC_{voxel}$ .

401 **Generalisation criterion.** Although generalisation is inherently continuous, we  
402 defined a practical criterion for it based on the expected performance under random  
403 chance. Methods with performance overlapping with or below the chance-level  
404 AUPRC were considered to have failed to generalise. The chance-level AUPRC value  
405 is equivalent to the prevalence of the positive class within a given region of interest  
406 (Saito and Rehmsmeier, 2015). In our case, this corresponds to the ratio between the  
407 PVS volume in the ground truth and the total volume of the region of interest, i.e., the  
408 fractional BG/CSO PVS volumes for each dataset (Table 1).

### 409 **3.3 Regions of interest**

410 We applied SynthSeg (Billot et al., 2023a) to T2w or T1w images to obtain  
411 parcellations, which we then aggregated to generate masks for the basal ganglia and

412 the centrum semiovale region of interest (BG ROI and CSO ROI). The BG ROIs  
413 included the internal and external capsules, caudate, lentiform, and thalamic nuclei,  
414 while the CSO ROI covered the remaining supratentorial white matter. While these  
415 two ROIs do not precisely match anatomical structures, we adhered to the established  
416 nomenclature to maintain consistency with widely used visual rating methods in the  
417 field (Potter et al., 2015). We refined these masks to guarantee the exclusion of the  
418 ventricular atrium, choroid plexus, and posterior horns of the lateral ventricles via atlas  
419 registration (<https://doi.org/10.7488/ds/1369>). All regions of interest were kept identical  
420 across evaluated methods to ensure that observed differences arose from the  
421 methods themselves rather than from variations in ROI definition.

## 422 **3.4 Competing methods**

423 We compared DRIPS against four other methods: the Frangi filter (Frangi et al., 1998),  
424 RORPO (Ranking the Orientation Responses of Path Operators) (Merveille et al.,  
425 2018, 2014), SHIVA-PVS (Boutinaud et al., 2021b), and nnU-Net (Pham et al., 2024).  
426 Both SHIVA-PVS and nnU-Net were used as pretrained models, tested only in an out-  
427 of-sample setting, with no training performed on the cohorts used in this study.

428 Frangi and RORPO are classical strategies designed for enhancing tubular structures.  
429 Frangi relies on Hessian-based voxel analysis of shape features, while RORPO  
430 applies multi-orientation path opening to distinguish tubular from spherical structures.  
431 We employed a thoroughly validated pipeline developed at the University of Edinburgh  
432 that integrates both methods (more details can be found in (Ballerini et al., 2018;  
433 Bernal et al., 2022; Duarte Coello et al., 2024; Valdés Hernández et al., 2024); the  
434 step-by-step pipeline can be found in <https://datashare.ed.ac.uk/handle/10283/8501>).  
435 Unlike standard Frangi filter implementations, the pipeline modifies the Gaussian

436 filtering step to handle anisotropic voxel sizes. We employed the uint8 conversion step  
437 for RORPO provided in the pipeline and used parameter settings derived from earlier  
438 optimisation studies (Frangi:  $\sigma_{\min} = 0.4$ ,  $\sigma_{\max} = 1.2$ ,  $\sigma_{\text{step}} = 0.2$ ,  $\alpha = 0.5$ ,  $\beta = 0.5$ , and  $c$   
439 = 500; RORPO: scaleMin=1, nbscales = 9, factor=1.7, dilationSize=1) (Ballerini et al.,  
440 2018; Bernal et al., 2022; Duarte Coello et al., 2024). We did not use any other pre-  
441 or post-processing strategies.

442 SHIVA-PVS is a U-Net-based convolutional neural network designed to segment PVS  
443 on T1w MRI scans. It requires input images of size  $160 \times 214 \times 176$ , with a  $1 \times 1 \times 1$   
444  $\text{mm}^3$  isotropic resolution and intensity values normalised to [0,1]. Pre-processing  
445 involved rigid registration of all T1w images to MNI space, cropping to the required  
446 dimensions, and applying min–max normalisation. Following inference, the resulting  
447 segmentations were padded and transformed back to native space using the inverse  
448 rigid registration. The algorithm requires no parameter tuning and is publicly available  
449 on GitHub: [https://github.com/pboutinaud/SHIVA\\_PVS](https://github.com/pboutinaud/SHIVA_PVS).

450 nnU-Net is a convolutional neural network that extends the no-new-U-Net (nnU-Net)  
451 (Isensee et al., 2021) for PVS segmentation. Two modality-specific models were  
452 trained, one for T1w and one for T2w images. We refer to the models as nnU-Net  
453 (T1w) and nnU-Net (T2w), respectively. Models requires no manual parameter tuning,  
454 as all pre-processing, processing, and post-processing steps are automated and  
455 implemented in the publicly available codebase: [https://github.com/wpham17/nnUNet-  
456 Perivascular-Spaces](https://github.com/wpham17/nnUNet-Perivascular-Spaces).

### 457 **3.5 Generalisation to other imaging modalities**

458 Since our aim was to assess the generalisation capabilities of models trained with  
459 DRIPS and the transferability of its learnt features, we also examined whether it could

460 extend to imaging modalities beyond MRI. As a proof of concept, we applied it to a 3D  
461 ex-vivo model of the human brain (Amunts et al., 2013). The Human Brain Histology  
462 dataset provides an ultrahigh-resolution 3D model of the human brain reconstructed  
463 from 7404 histological sections. For compatibility with our models and due to hardware  
464 constraints, the data were converted to greyscale and downsampled to 1 mm<sup>3</sup>  
465 resolution. Manual PVS segmentation was then performed on five axial slices in the  
466 BG ROI and five axial slices in the CSO ROI by an experienced image analyst using  
467 ITK-Snap with the smallest available drawing tools.

## 468 **4 Results**

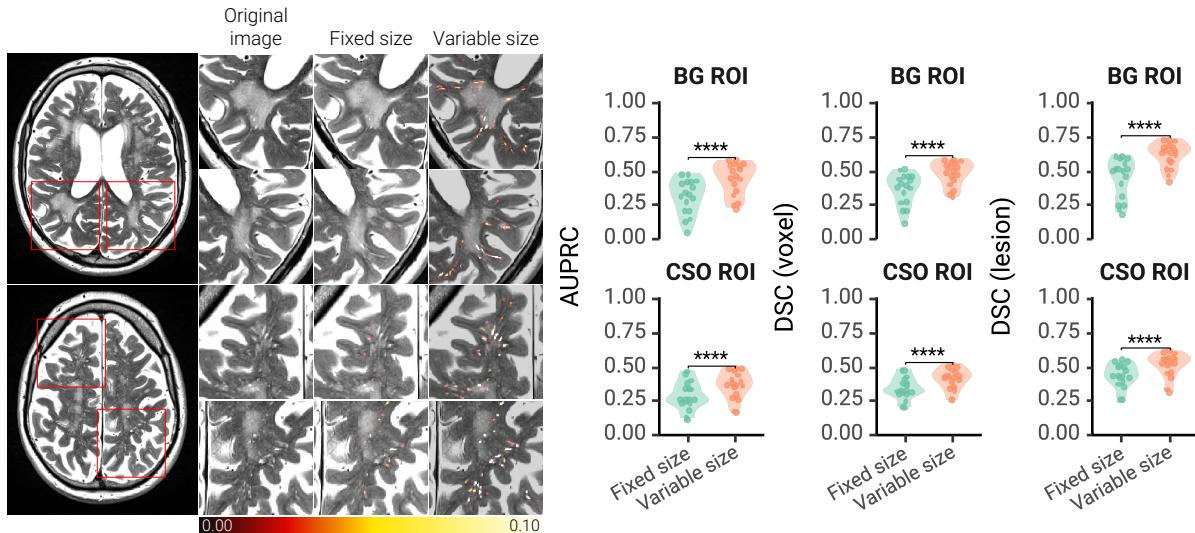
### 469 **4.1 Ablation study**

470 We evaluated the impact of individual DRIPS modules by comparing a model  
471 incorporating them with one that did not. We conducted these assessments on real  
472 data. It should be noted that the real data were not modified in any way.

#### 473 **4.1.1 Effect of voxel size variation in DRIPS on model performance**

474 To assess the effect of resampling and voxel size variation in DRIPS (Section 2.2.1),  
475 we compared the performance of two models: one with fixed and one with variable  
476 voxel sizes. We did this evaluation using data from the EBBIVD cohort (Figure 4). The  
477 use of variable voxel sizes led to a significant (P<0.001) and consistent improvement  
478 in segmentation performance. In the BG, median AUPRC improved from 0.325  
479 [0.210–0.423] to 0.459 [0.358–0.541], DSC<sub>voxel</sub> from 0.397 [0.276–0.459] to 0.499  
480 [0.432–0.555], and DSC<sub>lesion</sub> from 0.508 [0.322–0.552] to 0.635 [0.567–0.679]. In the  
481 CSO, median AUPRC rose from 0.256 [0.236–0.338] to 0.363 [0.304–0.439], DSC<sub>voxel</sub>

482 from 0.323 [0.305–0.386] to 0.423 [0.348–0.463], and  $DSC_{lesion}$  from 0.435 [0.393–  
483 0.506] to 0.532 [0.461–0.570].



484

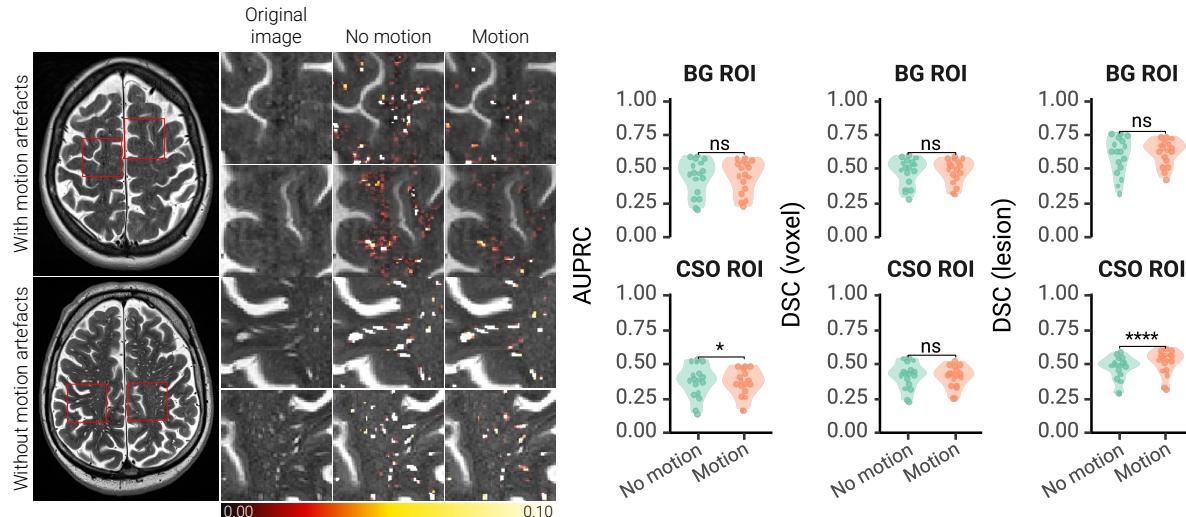
485 **Figure 4. Allowing variable voxel sizes during image generation yielded better segmentation**  
486 **performance than using fixed voxel sizes.** To illustrate this effect, we evaluated performance on  
487 EBBIVD, a cohort with highly anisotropic voxels (2.2 mm). Fewer PVS were segmented when models  
488 were trained with fixed voxel sizes compared to variable ones (left). With fixed voxel sizes, the model  
489 systematically missed multiple PVS in both normal-appearing white matter and WMH. For clarity,  
490 outputs were truncated to the 0.0–0.10 interval. At the cohort level, Wilcoxon signed-rank tests  
491 confirmed significant differences ( $P<0.0001$ ) in both AUPRC and DSC across regions of interest (right).

492

#### 493 **4.1.2 Effect of motion simulation in DRIPS on model performance**

494 We assessed how simulating motion in DRIPS influenced segmentation performance  
495 (Section 2.2.4), using data from the EBBIVD cohort. We compared the performance  
496 of two models: one incorporating motion simulation during training and one not (Figure  
497 4). Motion simulation enhanced the model's ability to distinguish true PVS from motion-  
498 induced ghosting, as illustrated in case-level examples with and without visible motion.  
499 At the group level, where both motion-affected and unaffected images are present,  
500 performance in the BG was comparable between models: AUPRC 0.469 [0.336–  
501 0.545] without vs. 0.459 [0.358–0.541] with motion,  $DSC_{voxel}$  0.509 [0.406–0.556] vs.  
502 0.499 [0.432–0.555], and  $DSC_{lesion}$  0.628 [0.517–0.701] vs. 0.635 [0.567–0.679]. In

503 the CSO, however, the motion-trained model was slightly more conservative voxel-  
504 wise, with AUPRC 0.390 [0.299–0.432] vs. 0.363 [0.304–0.439] ( $P=0.031$ ), but  
505 achieved a higher DSC<sub>lesion</sub>, 0.493 [0.430–0.510] vs. 0.532 [0.461–0.570] ( $P<0.001$ ).



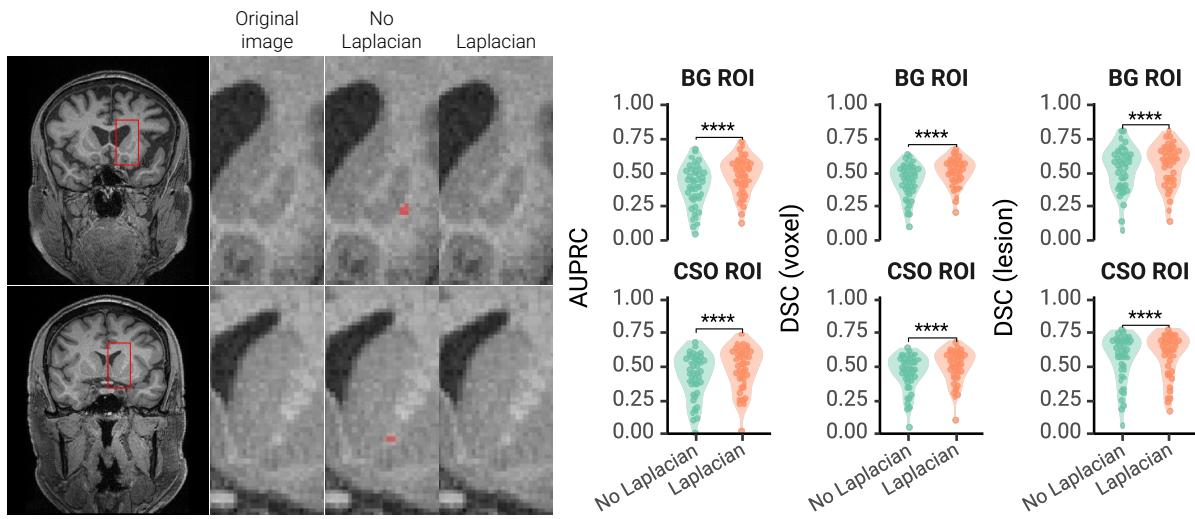
507 **Figure 5. Incorporating motion artefacts during image generation results in a more conservative**  
508 **model with improved ability to separate motion artefacts from PVS.** To illustrate this effect, we  
509 evaluated performance on EBBIVD and show probability maps for two cases: one with visible motion  
510 artefacts (left top row) and one without (left bottom row). When trained with motion artefacts, the model  
511 demonstrated improved ability to separate true PVS from motion-induced ghosting. In the motion case,  
512 the best DSC<sub>voxel</sub> and DSC<sub>lesion</sub> values without motion augmentation were 0.288 and 0.235, respectively,  
513 whereas training with motion artefacts increased them to 0.334 and 0.258. In cases without visible  
514 motion artefacts, both models yielded comparable results. At the group level, where images with and  
515 without motion artefacts are present, no differences were observed for BG PVS. For CSO PVS,  
516 however, the model trained with motion artefacts tended to be more conservative in detection but, once  
517 optimally thresholded, identified lesions better than the model trained without motion artefacts.

518

#### 519 **4.1.3 Effect of Laplacian constraint on model performance**

520 We tested whether applying a Laplacian constraint to restrict detections to hypointense  
521 structures on T1w images and hyperintense structures on T2w images improved  
522 segmentations yielded by DRIPS (Section 2.3.2). We compared the performance of  
523 DRIPS with and without post-processing of its outputs using data from PCB (Figure  
524 6). The Laplacian constraint significantly ( $P<0.0001$ ) reduced the number of false  
525 positives, leading to overall improvements in PVS segmentation. In the BG ROI,

526 AUPRC increased from 0.416 [0.307–0.506] to 0.494 [0.409–0.567],  $DSC_{voxel}$  from  
527 0.465 [0.373–0.528] to 0.509 [0.441–0.570], and  $DSC_{lesion}$  from 0.571 [0.406–0.635]  
528 to 0.590 [0.445–0.667]. In the CSO ROI, AUPRC improved from 0.454 [0.321–0.558]  
529 to 0.515 [0.380–0.611],  $DSC_{voxel}$  from 0.479 [0.397–0.555] to 0.522 [0.424–0.590], and  
530  $DSC_{lesion}$  from 0.615 [0.477–0.694] to 0.635 [0.471–0.692].



532 **Figure 6. With the Laplacian constraint, only tubular structures matching the expected intensity**  
533 **profiles are detected.** We assessed the effect of the Laplacian constraint on segmentation  
534 performance using the PCB cohort, which consists of T1w images where PVS appear hypointense.  
535 Because DRIPS is contrast-agnostic, it disregards intensity information. As a result, models trained with  
536 DRIPS may identify tubular structures regardless of whether they are hypo- or hyperintense, even  
537 though PVS present with a specific intensity profile. Such cases occurred most frequently within the  
538 internal and external capsules. By retaining regions with positive Laplacian values in T1w and negative  
539 values in T2w images, the Laplacian constraint reduced false positives and improved the quality of PVS  
540 segmentation overall.

## 541 4.2 Out-of-sample PVS segmentation

542 We compared DRIPS against Frangi, RORPO, SHIVA-PVS, and nnU-Net (Table 2).  
543 Below, we focus on two aspects: whether methods generalise out-of-sample and, if  
544 so, how they compare with one another.

#### 545 4.2.1 Generalisation

546 DRIPS generalised across all cohorts, independent of voxel anisotropy or image  
547 modality (T1w/T2w). The other method that ran successfully on all datasets was the  
548 Frangi filter. RORPO was not able to segment any PVS on ADNI. The generalisation  
549 of SHIVA-PVS and the nnU-Net models was limited to their respective training  
550 modalities, with AUPRC values overlapping with or falling below those of a random  
551 classifier when applied to unseen modalities.

#### 552 4.2.2 Segmentation performance

553 In cohorts with isotropic T1w imaging (PCB and ADNI), DRIPS and nnU-Net (T1w)  
554 were the top performers. Compared to the third-best method, they showed median  
555 improvements of +0.17–0.39 in AUPRC, +0.09–0.26 in  $DSC_{voxel}$ , and +0.14–0.25 in  
556  $DSC_{lesion}$ .

557 In PCB, DRIPS and nnU-Net (T1w) performed similarly in the BG ROI, with no  
558 significant differences across AUPRC (0.504 [0.406–0.568] vs 0.445 [0.421–0.535],  
559  $P=0.644$ ),  $DSC_{voxel}$  (0.512 [0.440–0.570] vs 0.516 [0.483–0.571],  $P=0.163$ ) or  $DSC_{lesion}$   
560 (0.589 [0.442–0.667] vs 0.553 [0.473–0.619],  $P=0.396$ ). In the CSO ROI, however,  
561 nnU-Net (T1w) achieved significantly higher scores, leaving DRIPS as the second-  
562 best performer (AUPRC: 0.515 [0.369–0.608] vs 0.549 [0.450–0.636],  $P<0.001$ ;  
563  $DSC_{voxel}$ : 0.521 [0.418–0.587] vs 0.543 [0.484–0.610],  $P<0.001$ ;  $DSC_{lesion}$ : 0.630  
564 [0.460–0.690] vs 0.649 [0.581–0.736],  $P<0.001$ ).

565 In ADNI, DRIPS significantly outperformed nnU-Net (T1w) in the BG ROI across all  
566 metrics (AUPRC: 0.569 [0.417–0.662] vs 0.474 [0.340–0.535],  $P=0.003$ ;  $DSC_{voxel}$ :  
567 0.564 [0.460–0.651] vs 0.322 [0.175–0.421],  $P<0.001$ ;  $DSC_{lesion}$ : 0.685 [0.571–0.823]  
568 vs 0.517 [0.437–0.588],  $P<0.001$ ). In the CSO ROI, results were more balanced:

569 DRIPS had higher sensitivity, detecting more PVS ( $DSC_{lesion}$ : 0.680 [0.616–0.720] vs  
570 0.593 [0.534–0.667],  $P<0.001$ ), while nnU-Net (T1w) provided slightly more precise  
571 delineation ( $DSC_{voxel}$ : 0.636 [0.575–0.657] vs 0.652 [0.563–0.690],  $P=0.040$ ).

572 In cohorts with anisotropic T2w imaging (EBBIVD, HIM, and MD-DARS), DRIPS had  
573 the best performance, followed generally by Frangi, RORPO, and the nnU-Net (T2w)  
574 in that order. The performance gap between DRIPS and the second-best method was  
575 most pronounced in the CSO ROI, with median gains of +0.17-0.22 in AUPRC, +0.12-  
576 0.14 in  $DSC_{voxel}$ , and +0.06-0.09 in  $DSC_{lesion}$ . In the BG ROI, the gap was smaller yet  
577 consistent, with AUPRC gains of +0.13-0.17,  $DSC_{voxel}$  gains of +0.07-0.12, and  
578  $DSC_{lesion}$  gains of +0.03-0.09.

579 SHIVA-PVS typically underperformed (AUPRC<0.10;  $DSC<0.15$ ), with the only  
580 exception in CSO PVS segmentation in ADNI, where it placed third above Frangi.

581 **Table 2. Out-of-sample PVS segmentation performance across five cohorts.** We assessed PVS segmentation in the basal ganglia (BG ROI) and the  
 582 centrum semiovale (CSO ROI) using voxel- and lesion-wise Dice similarity coefficients ( $DSC_{voxel}$  and  $DSC_{lesion}$ ) and the area under the precision–recall curve  
 583 (AUPRC). We report medians with interquartile ranges, and “NA” where no PVS could be segmented. We identified the best-performing methods across regions  
 584 and cohorts using the Wilcoxon signed-rank test and highlighted them in bold. Following the generalisability criterion described in Section 3.2, we marked with  
 585 “NG” all AUPRC values that overlapped with or fell below the expected performance of a random classifier.

	Method	Metric	PCB (N=42)	EBBIVD (N=18)	HIM (N=39)	MD-DARS (N=6)	ADNI (N=60)
BG ROI	Frangi	AUPRC	0.263 [0.213–0.321]	0.334 [0.272–0.348]	0.331 [0.269–0.360]	0.294 [0.253–0.314]	0.198 [0.101–0.324]
		$DSC_{voxel}$	0.331 [0.292–0.391]	0.425 [0.379–0.439]	0.407 [0.372–0.443]	0.388 [0.345–0.404]	0.342 [0.247–0.445]
		$DSC_{lesion}$	0.447 [0.390–0.493]	0.592 [0.524–0.629]	0.556 [0.516–0.639]	0.556 [0.529–0.596]	0.467 [0.323–0.583]
	RORPO	AUPRC	0.203 [0.168–0.287]	0.324 [0.234–0.351]	0.308 [0.214–0.361]	0.264 [0.219–0.336]	NA
		$DSC_{voxel}$	0.303 [0.242–0.374]	0.417 [0.376–0.453]	0.415 [0.356–0.451]	0.381 [0.347–0.421]	NA
		$DSC_{lesion}$	0.252 [0.207–0.326]	0.424 [0.376–0.541]	0.432 [0.363–0.493]	0.421 [0.365–0.450]	NA
	SHIVA-PVS	AUPRC	0.060 [0.049–0.073]	0.018 [0.015–0.025] (NG)	0.013 [0.010–0.018] (NG)	0.013 [0.011–0.019] (NG)	0.101 [0.037–0.158]
		$DSC_{voxel}$	0.134 [0.114–0.154]	0.040 [0.032–0.048]	0.032 [0.028–0.039]	0.036 [0.033–0.042]	0.214 [0.108–0.285]
		$DSC_{lesion}$	0.242 [0.193–0.275]	0.039 [0.023–0.094]	0.078 [0.039–0.103]	0.094 [0.062–0.108]	0.426 [0.330–0.520]
CSO ROI	nn-Unet (T1w)	AUPRC	<b>0.445 [0.421–0.535]</b>	0.009 [0.006–0.012] (NG)	0.006 [0.005–0.008] (NG)	0.007 [0.004–0.008] (NG)	0.474 [0.340–0.535]
		$DSC_{voxel}$	<b>0.516 [0.483–0.571]</b>	0.031 [0.024–0.045]	0.021 [0.017–0.032]	0.026 [0.018–0.033]	0.322 [0.173–0.421]
		$DSC_{lesion}$	<b>0.553 [0.473–0.619]</b>	0.028 [0.023–0.035]	0.032 [0.027–0.041]	0.035 [0.025–0.044]	0.517 [0.437–0.588]
	nn-Unet (T2w)	AUPRC	0.009 [0.006–0.011] (NG)	0.100 [0.057–0.141]	0.107 [0.082–0.136]	0.087 [0.078–0.106]	0.002 [0.001–0.006] (NG)
		$DSC_{voxel}$	0.027 [0.022–0.037]	0.208 [0.121–0.251]	0.225 [0.173–0.254]	0.216 [0.180–0.220]	0.013 [0.005–0.027]
		$DSC_{lesion}$	0.050 [0.042–0.065]	0.278 [0.184–0.344]	0.282 [0.217–0.343]	0.283 [0.202–0.365]	0.041 [0.023–0.090]
	DRIPS	AUPRC	<b>0.504 [0.406–0.568]</b>	<b>0.459 [0.358–0.541]</b>	<b>0.503 [0.426–0.553]</b>	<b>0.424 [0.396–0.447]</b>	<b>0.569 [0.417–0.662]</b>
		$DSC_{voxel}$	<b>0.512 [0.440–0.570]</b>	<b>0.499 [0.432–0.555]</b>	<b>0.532 [0.488–0.567]</b>	<b>0.475 [0.471–0.478]</b>	<b>0.564 [0.460–0.651]</b>
		$DSC_{lesion}$	<b>0.589 [0.442–0.667]</b>	<b>0.635 [0.567–0.679]</b>	<b>0.646 [0.618–0.697]</b>	<b>0.581 [0.571–0.478]</b>	<b>0.685 [0.571–0.823]</b>
CSO ROI	Frangi	AUPRC	0.170 [0.119–0.276]	0.185 [0.129–0.253]	0.192 [0.156–0.250]	0.150 [0.121–0.180]	0.272 [0.157–0.362]
		$DSC_{voxel}$	0.311 [0.249–0.409]	0.286 [0.249–0.341]	0.314 [0.268–0.340]	0.260 [0.238–0.289]	0.381 [0.269–0.447]
		$DSC_{lesion}$	0.429 [0.333–0.546]	0.472 [0.383–0.515]	0.451 [0.394–0.514]	0.391 [0.340–0.440]	0.426 [0.362–0.509]
	RORPO	AUPRC	0.181 [0.103–0.309]	0.196 [0.119–0.244]	0.181 [0.127–0.263]	0.148 [0.112–0.179]	NA
		$DSC_{voxel}$	0.311 [0.219–0.429]	0.301 [0.220–0.354]	0.290 [0.227–0.360]	0.253 [0.208–0.287]	NA
		$DSC_{lesion}$	0.376 [0.252–0.484]	0.344 [0.267–0.408]	0.309 [0.264–0.388]	0.298 [0.217–0.325]	NA
	SHIVA-PVS	AUPRC	0.011 [0.005–0.022]	0.026 [0.018–0.032] (NG)	0.013 [0.010–0.018] (NG)	0.019 [0.013–0.020] (NG)	0.421 [0.319–0.481]
		$DSC_{voxel}$	0.035 [0.019–0.056]	0.049 [0.035–0.060]	0.039 [0.029–0.049]	0.038 [0.025–0.042]	0.469 [0.417–0.521]
		$DSC_{lesion}$	0.063 [0.038–0.110]	0.014 [0.001, 0.031]	0.014 [0.005, 0.023]	0.010 [0.005–0.018]	0.564 [0.487–0.652]
CSO ROI	nn-Unet (T1w)	AUPRC	<b>0.549 [0.450–0.636]</b>	0.013 [0.008–0.016] (NG)	0.010 [0.007–0.012] (NG)	0.009 [0.007–0.011] (NG)	<b>0.632 [0.564–0.697]</b>
		$DSC_{voxel}$	<b>0.543 [0.484–0.610]</b>	0.047 [0.029–0.059]	0.037 [0.028–0.047]	0.037 [0.025–0.040]	<b>0.652 [0.563–0.690]</b>
		$DSC_{lesion}$	<b>0.649 [0.581–0.736]</b>	0.010 [0.009, 0.014]	0.010 [0.008–0.015]	0.013 [0.007–0.015]	0.593 [0.534–0.667]
	nn-Unet (T2w)	AUPRC	0.003 [0.017–0.005] (NG)	0.140 [0.106–0.177]	0.160 [0.124–0.206]	0.116 [0.105–0.136]	0.007 [0.004–0.010] (NG)
		$DSC_{voxel}$	0.007 [0.005–0.012]	0.216 [0.181–0.246]	0.256 [0.210–0.302]	0.209 [0.200–0.224]	0.019 [0.013–0.029]
		$DSC_{lesion}$	0.014 [0.010–0.019]	0.296 [0.263–0.327]	0.348 [0.293–0.399]	0.284 [0.232–0.330]	0.046 [0.028–0.059]
	DRIPS	AUPRC	0.515 [0.369–0.608]	<b>0.363 [0.304–0.439]</b>	<b>0.409 [0.336–0.464]</b>	<b>0.323 [0.286–0.358]</b>	<b>0.665 [0.569–0.698]</b>
		$DSC_{voxel}$	0.521 [0.418–0.587]	<b>0.423 [0.348–0.463]</b>	<b>0.452 [0.399–0.482]</b>	<b>0.387 [0.352–0.412]</b>	0.636 [0.575–0.657]
		$DSC_{lesion}$	0.630 [0.460–0.690]	<b>0.532 [0.461–0.570]</b>	<b>0.545 [0.498–0.616]</b>	<b>0.467 [0.402–0.521]</b>	<b>0.680 [0.616–0.720]</b>

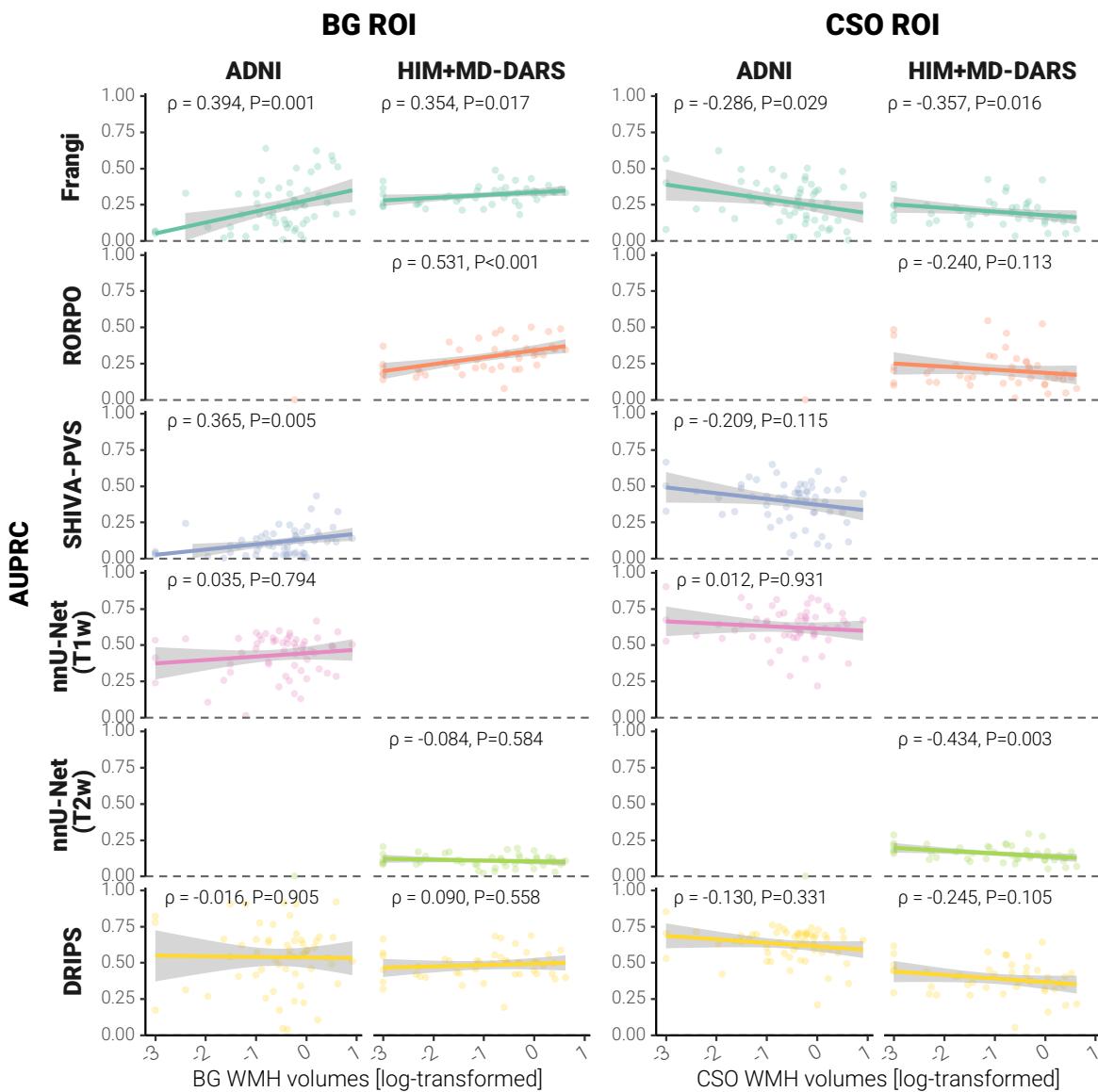
### 587 4.2.3 WMH and PVS segmentation

588 WMH can impair accurate PVS segmentation. To assess this effect, we examined the  
589 relationship between WMH volume and AUPRC using Spearman correlations (Figure  
590 7). For this secondary analysis, we used data from ADNI, HIM, and MD-DARS (N =  
591 105), all of which had WMH segmentations. We combined HIM and MD-DARS due to  
592 the small sample size of MD-DARS, which could otherwise lead to spurious  
593 correlations. The analysis focused on models that successfully generalised.

594 In the BG ROI, AUPRC values obtained by the Frangi filter in both T1w and T2w  
595 imaging, by RORPO in T2w imaging, and by SHIVA-PVS in T1w imaging increased  
596 with greater WMH volume ( $P \leq 0.01$ ). The underlying reasons differed between  
597 SHIVA-PVS and the Frangi filter or RORPO. SHIVA-PVS performed better in cases  
598 with more visible BG PVS (Figure 8), which occurred more frequently in patients with  
599 greater WMH burden (Spearman correlation between BG PVS volume and BG WMH  
600 volume in ADNI:  $\rho = 0.396$ ,  $P = 0.002$ ). Both the Frangi filter and RORPO produced  
601 non-zero responses within WMH. In patients with higher BG WMH burden, many WMH  
602 voxels were adjacent or around to true PVS, causing false detections to overlap with  
603 true positives and artificially inflating recall rates and AUPRC (Figure 8). Unlike Frangi,  
604 RORPO, and SHIVA-PVS, AUPRC of the nnU-Net models (T1w and T2w) and DRIPS  
605 in the BG ROI did not relate to BG WMH volumes ( $P > 0.10$ ).

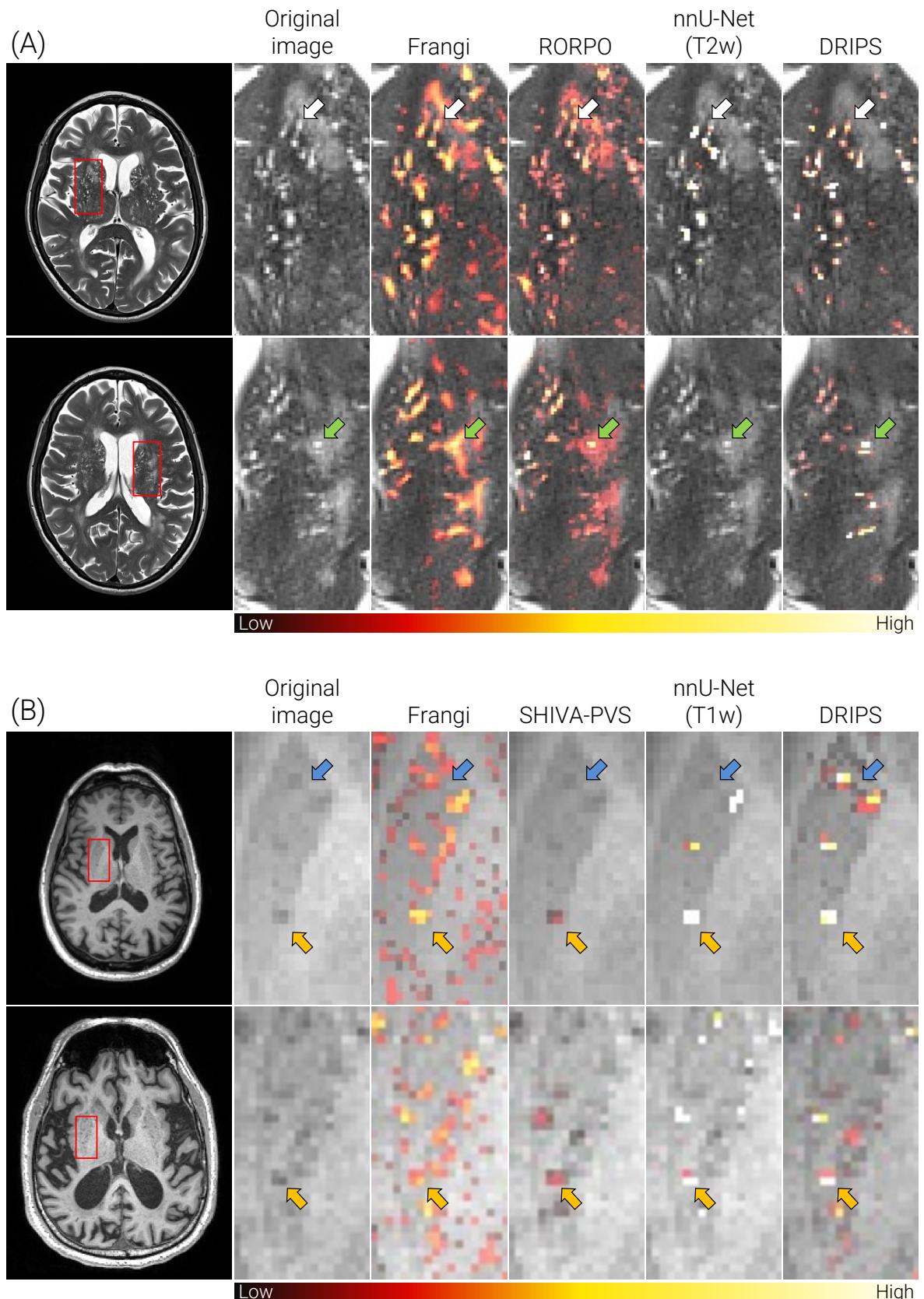
606 In the CSO ROI, AUPRC values of the Frangi filter in both T1w and T2w imaging, as  
607 well as that of the nnU-Net (T2w) in T2w imaging, declined with increasing WMH  
608 volume in the same region ( $P < 0.05$ ). The reasons behind these associations differed  
609 between methods. The Frangi filter generally marked WMH as potential PVS. As a  
610 result, higher WMH burden produced more false positives and consequently lower

611 AUPRC values (Figure 8). In contrast, the nnU-Net (T2w) more effectively disregarded  
 612 WMH as potential PVS candidates, but this same ability also led to the omission of  
 613 PVS located within WMH regions (Figure 8). The AUPRC values obtained by RORPO,  
 614 SHIVA-PVS, nnU-Net (T1w), and DRIPS in the CSO ROI were not associated with  
 615 CSO WMH volumes ( $P > 0.05$ ).



616

617 **Figure 7. Relationship between segmentation performance (AUPRC) and regional WMH volume**  
 618 **for each algorithm across the ADNI and HIM and MD-DARS.** We studied these relationships using  
 619 Spearman correlation coefficients (shown above each subplot). Algorithms that failed or showed limited  
 620 generalisation within specific datasets were excluded from this secondary analysis (ADNI: RORPO,  
 621 nnU-Net (T2w); HIM+MD-DARS: SHIVA-PVS, nnU-Net (T1w)). We used the Greek letter  $\rho$  to denote  
 622 the Spearman correlation coefficient and  $P$  to denote its p-value.



623

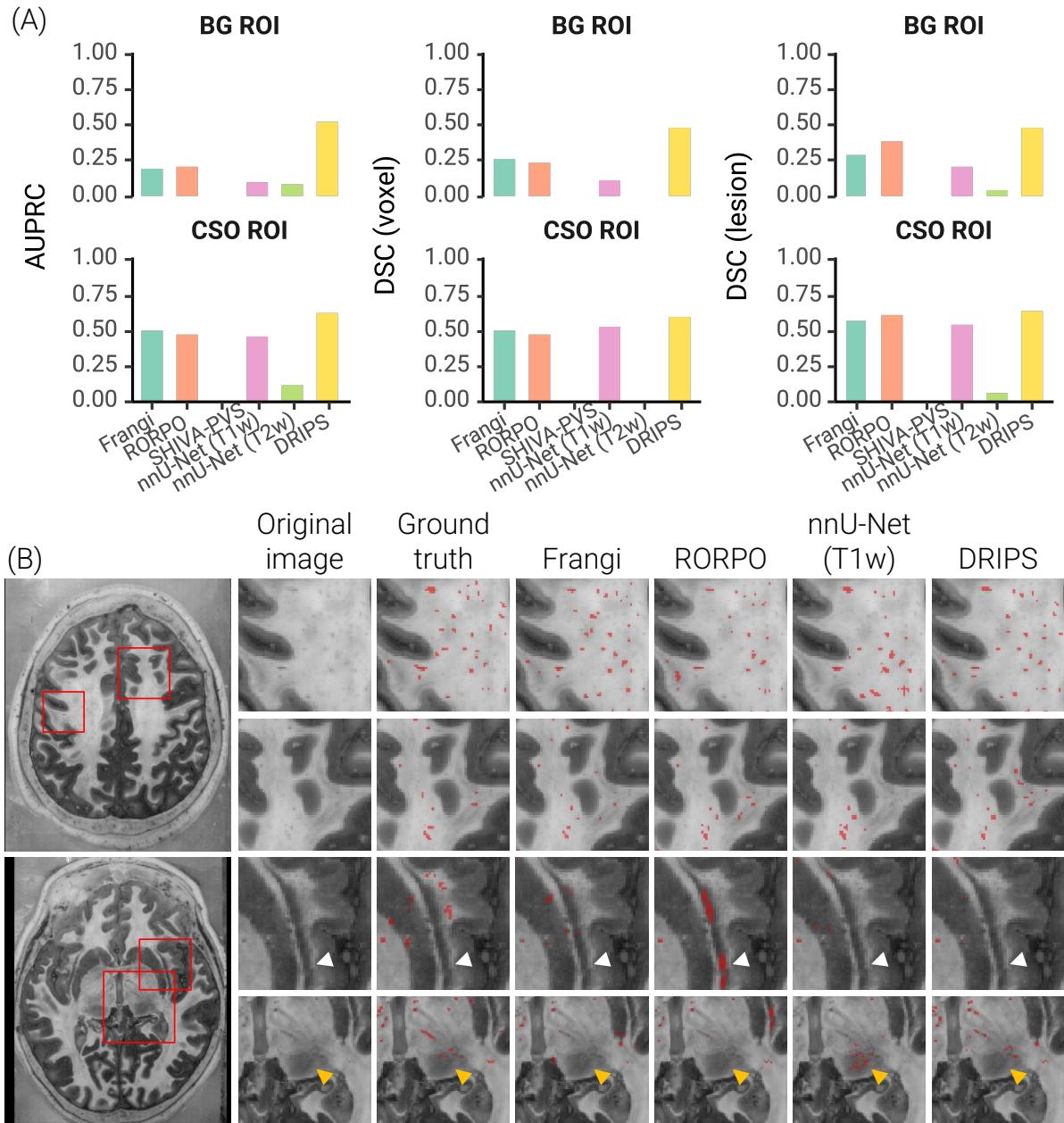
624 **Figure 8. Response map yielded by PVS segmentation methods on T2w and T1 imaging.** (A)  
625 Maps obtained from all methods that generalised to T2w imaging. The Frangi filter and RORPO  
626 produced non-zero responses within WMH. In patients with higher BG WMH burden, false detections

627 near true PVS often overlapped spatially, artificially inflating AUPRC (white and green arrows). On the  
628 other hand, the nnU-Net (T2w) tended to miss PVS located within WMH (green arrow). (B) Maps  
629 obtained from all methods that generalised to T1w imaging. SHIVA-PVS identified salient as opposed  
630 to subtle PVS (yellow vs blue arrows).

### 631 **4.3 Generalisation to other imaging modalities**

632 We evaluated the generalisation capacity of DRIPS and the four competing methods  
633 beyond MRI, with particular emphasis on their transferability to a 3D *ex vivo* brain  
634 model reconstructed from histology (Figure 9). Histology-to-MNI registration was  
635 unsuccessful with SynthMorph and ANTs, preventing SHIVA-PVS from being  
636 evaluated. DRIPS achieved the best performance across both BG and CSO ROIs. In  
637 the BG, it reached a  $DSC_{lesion}$  of 0.477,  $DSC_{voxel}$  of 0.482, and AUPRC of 0.512, clearly  
638 outperforming all other methods (next best  $DSC_{lesion}$  0.373 with RORPO,  $DSC_{voxel}$   
639 0.260 with Frangi, and AUPRC 0.205 with RORPO). In the CSO, it again obtained the  
640 highest scores with  $DSC_{lesion}$  0.629,  $DSC_{voxel}$  0.592, and AUPRC 0.625, surpassing  
641 RORPO (0.607/0.466/0.475), nnU-Net (T1w; 0.542/0.517/0.450), and Frangi  
642 (0.564/0.492/0.493). nnU-Net (T2w) did not generate meaningful PVS segmentations.

643



644

645 **Figure 9. PVS segmentation on a 3D ex-vivo brain model reconstructed from histology images.**  
646 (A) Segmentation performance, as measured by AUPRC and Dice at both voxel and lesion level.  
647 Registration of histology images to MNI space was unsuccessful with SynthMorph and ANTs,  
648 preventing SHIVA-PVS evaluation. The nnU-Net (T2w) could not segment PVS successfully. The  
649 classical methods, Frangi and RORPO, successfully segmented PVS as expected due to their modality-  
650 agnostic design. Both nnU-Net (T1w) and DRIPS produced valid segmentations. DRIPS outperformed  
651 all other methods across both regions of interest. RORPO and Frangi achieved the next best results,  
652 while nnU-Net (T1w) had the lowest performance. (B) Example segmentations in the CSO and BG ROIs  
653 of histology sections. Across algorithms, thresholds were chosen to yield the highest voxel-level Dice  
654 coefficients. Segmentation in the CSO ROI was successful across methods, whereas performance in  
655 the BG ROI was impaired by systematic errors, including misclassification of the claustrum as PVS  
656 (white arrow) and spurious segmentation of multiple thalamic structures as PVS (yellow arrow).

657 **5 Discussion**

658 In recent years, deep learning has become the dominant paradigm for PVS  
659 segmentation (Waymont et al., 2024) and for medical image analysis more broadly,  
660 primarily owing to its strong within-dataset performance. Yet, as also illustrated by our  
661 findings, such models often struggle to generalise when faced with data distributions  
662 or imaging modalities not represented during training. Unlike classical approaches—  
663 which may be less accurate in noisy settings but still provide a usable output—deep  
664 learning models can fail outright when applied outside their training domain, producing  
665 no meaningful segmentation. However, relying on constant manual labelling and fine-  
666 tuning for every new dataset is neither scalable nor sustainable.

667 Against this backdrop, our aim was not to develop a model narrowly optimised for a  
668 single dataset, but to propose a new PVS segmentation method that achieves high  
669 accuracy and robust generalisation across imaging sequences and cohorts. By  
670 leveraging physics-based image generation and domain randomisation, we  
671 demonstrated that it is possible to mitigate domain shifts and achieve accurate PVS  
672 segmentation under conditions seen during training. Across five independent cohorts,  
673 we show that DRIPS can (i) segment PVS on both isotropic and anisotropic T1- and  
674 T2w images, (ii) outperform classical and machine learning-based approaches, (iii)  
675 segment PVS independently of the overall WMH burden, and (iv) generalise even to  
676 other modalities, including histology. Taken together, these results position DRIPS as  
677 a robust and versatile framework for PVS segmentation.

678 **5.1 Physics-inspired domain randomisation**

679 DRIPS brings together two complementary research directions: domain randomisation  
680 and physics-inspired data augmentation. Domain randomisation tackles the challenge  
681 of generalisation by exposing models to synthetic data generated from segmentations  
682 with fully randomised parameters (Tobin et al., 2017), enabling the learning of robust  
683 and transferable features provided that the synthetic variability adequately reflects  
684 real-world conditions (Billot et al., 2023a). Physics-inspired data augmentation builds  
685 on this by modelling the image acquisition process and its artefacts, thereby enhancing  
686 realism and surpassing purely agnostic randomisation strategies (Adams et al., 2024).  
687 We observed that introducing voxel size variability through resampling and simulating  
688 motion artefacts both contributed positively to performance. Resampling proved  
689 essential for handling anisotropic scans. Overall, DRIPS achieved consistent  
690 improvements of approximately 0.10–0.13 across all evaluation metrics, with slightly  
691 greater gains in the BG compared to the CSO, when resampling was considered as  
692 opposed to when it was not. Simulating motion artefacts also helped models trained  
693 with DRIPS distinguish true PVS from motion-related ghosting, as seen in case-level  
694 examples. At the group level, performance in the BG was largely unaffected by motion  
695 training, whereas in the CSO it led to slightly lower voxel-wise precision–recall but  
696 significantly improved lesion-wise detection, suggesting a more conservative yet  
697 accurate segmentation strategy.

698 **5.2 DRIPS segments PVS accurately on real MRI data**

699 Conventional deep learning approaches to PVS segmentation have typically  
700 depended on small, carefully curated training datasets. While such models can  
701 achieve high accuracy within their training domain, they often fail to generalise well to

702 new datasets. SHIVA-PVS, a 3D U-Net trained solely on T1w images, exemplifies this  
703 limitation: it did not transfer to T2w scans and showed only limited sensitivity to PVS  
704 even within its training modality. A similar limitation was seen with the nnU-Net  
705 framework, where models trained on T1w images could only process T1w data, and  
706 likewise for T2w images, with little to no generalisation across modalities. Clearly,  
707 training separate models for each input modality offers a practical workaround, but it  
708 bypasses rather than addresses the fundamental issue of generalisability.

709 In contrast, our results highlight the utility of domain randomisation for bridging the  
710 generalisation gap. DRIPS had stable performance across both T1- and T2w images  
711 without the need for retraining, and importantly, the learned features also transferred  
712 to histological data—a modality entirely distinct from MRI. These findings reinforce the  
713 central premise of domain randomisation: that exposure to sufficiently diverse  
714 synthetic variation enables models to acquire representations that remain applicable  
715 beyond their original training domain.

### 716 **5.3 DRIPS versus competing approaches**

717 We compared DRIPS to both classical image-processing-based and machine  
718 learning-based methods, using scans and manual annotations from five cohorts ( $n =$   
719 165) that included healthy controls as well as individuals with Long-COVID,  
720 hypertensive arteriopathy, cerebral amyloid angiopathy, heart failure, mild cognitive  
721 impairment, and Alzheimer’s disease. DRIPS outperformed all competing methods on  
722 anisotropic scans (EBBIVD, HIM, and MD-DARS) and ranked among the top two on  
723 isotropic scans (PCB and ADNI).

724 On anisotropic T2w scans, the conventional Frangi filter generally emerged as the  
725 second-best method. This finding carries important implications for prior studies: when

726 carefully tuned, Frangi can achieve accurate PVS segmentation, outperforming all  
727 machine learning–based methods aside from DRIPS. Its main drawback, as with any  
728 other classical PVS segmentation strategy, is the need for manual calibration on each  
729 new dataset to reach optimal performance.

730 On isotropic T1w scans, nnU-Net and DRIPS achieved the highest overall  
731 performance, surpassing all other methods by median margins of at least 0.17 in  
732 AUPRC, 0.09 in  $DSC_{voxel}$ , and 0.14 in  $DSC_{lesion}$ . The marked improvement in precision  
733 and recall over classical image-processing methods likely stems from the fact that, as  
734 shown in Figure 8 and Figure 9, regions such as the boundaries of the putamen,  
735 pallidum, and claustrum are often misidentified as PVS by these methods solely due  
736 to their “tubular” appearance. Note that, in general, signal intensity differences  
737 between the basal ganglia and the surrounding white matter on T1w imaging—  
738 particularly at higher field strengths—can also be erroneously highlighted as PVS. In  
739 these situations, post-processing strategies that analyse jointly location, length and  
740 shape become essential. Their impact on the segmentation performance of classical  
741 techniques was not evaluated in this study, as it lay outside the primary scope of our  
742 work.

## 743 **5.4 Robustness against WMH**

744 Previous studies have shown that the presence of WMH can substantially compromise  
745 the performance of PVS segmentation methods (Bernal et al., 2022; Pham et al., 2022;  
746 Valdes Hernandez et al., 2013; Waymont et al., 2024). Our findings align with this  
747 evidence, revealing that both classical and deep learning approaches are often  
748 dependent on the regional WMH burden. Traditionally, one of the most common ways  
749 to mitigate this issue has been to exclude WMH from analyses. However, as illustrated

750 by the nnU-Net (T2w), this approach introduces its own bias: by excluding WMH, the  
751 method inherently omits PVS that overlap with them, creating artificial correlations with  
752 WMH volume, since individuals with more WMH also tend to have more PVS within  
753 them.

754 Both extremes—erroneously labelling WMH as PVS or excluding WMH entirely—are  
755 suboptimal. The goal should instead be to develop models whose performance is  
756 independent of WMH burden. In this regard, our results indicate that DRIPS was able  
757 to segment PVS comparably accurately even in cases with high WMH volumes,  
758 without its performance being significantly compromised or biased, regardless of  
759 whether input data were T1w or T2w images. A similar pattern was observed for the  
760 nnU-Net (T1w) in T1w imaging. Although these findings are based on a limited sample  
761 (60 T1w and 45 T2w images), they represent a promising step towards developing  
762 segmentation methods that are more robust and less biased by co-occurring brain  
763 lesions.

## 764 **5.5 Limitations and future work**

765 Despite the demonstrated generalisability of our approach, four limitations merit  
766 consideration. First, we modelled PVS as tortuous tubular structures distributed  
767 throughout the brain. While effective for training and segmentation, this abstraction  
768 oversimplifies their biology. *In vivo*, PVS are closely aligned with the cerebral  
769 vasculature, following the trajectories of arterioles, capillaries, and venules, with their  
770 orientation, calibre, and spatial density shaped by vascular anatomy, regional blood  
771 supply, and vessel tortuosity. Second, we assumed a predominant orientation towards  
772 the lateral ventricles. This is a reasonable approximation for PVS in the centrum  
773 semiovale, which often follow medullary arteries radiating to the ventricles, but it does

774 not hold in other regions—for instance, PVS surrounding the lenticulostriate arteries  
775 in the basal ganglia, which typically run perpendicular to the axial plane. Looking  
776 ahead, these limitations highlight an opportunity: conditioning PVS generation on  
777 vascular maps could produce more physiologically plausible simulations, improving  
778 anatomical fidelity and reducing false positives in regions where tubular structures  
779 occur independently of vessels. Third, in this work, we implemented the segmentation  
780 network in DRIPS as a 3D U-Net and did not investigate alternative, more advanced  
781 architectures. While this represents a limitation, it was not the primary focus of our  
782 study. Our main objective was to demonstrate that DRIPS can achieve accurate and  
783 robust PVS segmentation across multiple cohorts, health conditions, and imaging  
784 settings, rather than to develop a new method optimised for peak performance.  
785 Exploring whether more sophisticated segmentations models—such as nnU-Net or  
786 transformers—can further improve performance represents an important direction for  
787 future work. Fourth, although our evaluation included data from multiple individuals  
788 across five cohorts spanning a wide range of conditions—from normal cognition to  
789 post-COVID syndrome, hypertensive arteriopathy, cerebral amyloid angiopathy, heart  
790 failure, mild cognitive impairment, and Alzheimer’s disease—our assessment remains  
791 limited by the imaging protocols included in this study. As part of our future work, we  
792 plan to include patients spanning a broader range of disease severities—from very  
793 mild to advanced—scanned using multiple imaging sequences, and to conduct  
794 longitudinal assessments to evaluate the method’s ability to track changes in PVS over  
795 extended periods of time.

796 **6 Conclusion**

797 We introduced DRIPS, the first physics-inspired domain randomisation framework for  
798 accurate out-of-sample PVS segmentation. DRIPS accurately segmented PVS in both  
799 T1w and T2w images, at isotropic and anisotropic resolutions, without requiring  
800 manual PVS segmentations, retraining, or fine-tuning. It outperformed all competing  
801 methods on anisotropic images and achieved performance comparable to nnU-Net on  
802 isotropic data. Unlike the segmentation performance of competing methods, its  
803 performance was not associated by the volume of WMH in the brain. DRIPS's out-of-  
804 sample capabilities extended beyond MRI, successfully segmenting PVS in 3D *ex vivo*  
805 brain models reconstructed from histology. Collectively, our findings demonstrate that  
806 DRIPS segments PVS accurately across diverse imaging settings and patient  
807 populations, enabling more accessible and reliable automated PVS quantification for  
808 both research and clinical use.

809 **Declarations**

810 **Use of AI-assisted technologies in the manuscript**  
811 **preparation process**

812 The authors used ChatGPT to assist with grammar correction during the preparation  
813 of this work. All content was in all instances reviewed and edited by the authors, who  
814 take full responsibility for the final published article.

815 **Ethics approval and consent to participate**

816 The authors assert that all procedures contributing to this work comply with the ethical  
817 standards of the relevant national and institutional committees on human  
818 experimentation and with the Helsinki Declaration of 1975, as revised in 2008.

819 **Consent for publication**

820 Not applicable.

821 **Availability of data and materials**

822 We provide a ready-to-use Docker image for running DRIPS, available at  
823 <https://github.com/CIR-FAU/DRIPS.git>, to ensure easy deployment, reproducibility,  
824 and consistent performance across systems.

825 The datasets used and analysed during the current study are available from the  
826 corresponding author on reasonable request.

827 **Competing interests**

828 The authors report no competing interests.

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## 870 **CRediT authorship contribution statement**

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875 S.S., E.D., G.Z., J.B. Data Curation: L.B., J.B.; Writing - Original Draft: L.B., J.B.;  
876 Writing - Review & Editing: All authors; Visualisation: L.B., J.B.; Supervision: G.Z., J.B.;  
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## 878 **References**

879 Adams, R., Huynh, K.M., Zhao, W., Hu, S., Lyu, W., Ahmad, S., Ma, D., Yap, P.-T.,  
880 2024. UltimateSynth: MRI Physics for Pan-Contrast AI. bioRxiv 1–31.  
881 <https://doi.org/10.1101/2024.12.05.627056>

882 Amunts, K., Lepage, C., Borgeat, L., Mohlberg, H., Dickscheid, T., Rousseau, M.-É.,  
883 Bludau, S., Bazin, P.-L., Lewis, L.B., Oros-Peusquens, A.-M., Shah, N.J., Lippert,  
884 T., Zilles, K., Evans, A.C., 2013. BigBrain: An Ultrahigh-Resolution 3D Human  
885 Brain Model. *Science* (1979) 340. <https://doi.org/10.1126/science.1235381>

886 Ballerini, L., Lovreglio, R., Valdés Hernández, M.D.C., Ramirez, J., MacIntosh, B.J.,  
887 Black, S.E., Wardlaw, J.M., 2018. Perivascular Spaces Segmentation in Brain  
888 MRI Using Optimal 3D Filtering. *Sci Rep* 8, 1–11. <https://doi.org/10.1038/s41598-018-19781-5>

890 Ballerini, L., McGrory, S., Valdés Hernández, M. del C., Lovreglio, R., Pellegrini, E.,  
891 MacGillivray, T., Muñoz Maniega, S., Henderson, R., Taylor, A., Bastin, M.E.,  
892 Doubal, F., Trucco, E., Deary, I.J., Wardlaw, J., 2020. Quantitative measurements  
893 of enlarged perivascular spaces in the brain are associated with retinal  
894 microvascular parameters in older community-dwelling subjects. *Cereb Circ Cogn  
Behav* 1, 100002. <https://doi.org/10.1016/j.cccb.2020.100002>

896 Barisano, G., Iv, M., Choupan, J., Hayden-Gephart, M., 2025. Robust, fully-automated  
897 assessment of cerebral perivascular spaces and white matter lesions: a  
898 multicentre MRI longitudinal study of their evolution and association with risk of  
899 dementia and accelerated brain atrophy. *EBioMedicine* 111.  
900 <https://doi.org/10.1016/j.ebiom.2024.105523>

901 Barnes, A., Ballerini, L., Valdés Hernández, M. del C., Chappell, F.M., Muñoz  
902 Maniega, S., Meijboom, R., Backhouse, E. V., Stringer, M.S., Duarte Coello, R.,  
903 Brown, R., Bastin, M.E., Cox, S.R., Deary, I.J., Wardlaw, J.M., 2022. Topological  
904 relationships between perivascular spaces and progression of white matter

905 hyperintensities: A pilot study in a sample of the Lothian Birth Cohort 1936. *Front*  
906 *Neurol* 13. <https://doi.org/10.3389/fneur.2022.889884>

907 Bernal, J., Valdés-Hernández, M., Ballerini, L., Escudero, J., Jochems, A.C.C., Clancy,  
908 U., Doubal, F.N., Stringer, M.S., Thriplleton, M.J., Touyz, R.M., Wardlaw, J.M.,  
909 2020. A Framework for Jointly Assessing and Reducing Imaging Artefacts  
910 Automatically Using Texture Analysis and Total Variation Optimisation for  
911 Improving Perivascular Spaces Quantification in Brain Magnetic Resonance  
912 Imaging. *Communications in Computer and Information Science* 1248 CCIS,  
913 171–183. [https://doi.org/10.1007/978-3-030-52791-4\\_14](https://doi.org/10.1007/978-3-030-52791-4_14)

914 Bernal, J., Valdés-Hernández, M. d. C., Escudero, J., Heye, A.K., Sakka, E., Armitage,  
915 P.A., Makin, S., Touyz, R.M., Wardlaw, J.M., Thriplleton, M.J., 2021a. A four-  
916 dimensional computational model of dynamic contrast-enhanced magnetic  
917 resonance imaging measurement of subtle blood-brain barrier leakage.  
918 *Neuroimage* 230, 117786. <https://doi.org/10.1016/j.neuroimage.2021.117786>

919 Bernal, J., Valdés-Hernández, M.D.C., Escudero, J., Duarte, R., Ballerini, L., Bastin,  
920 M.E., Deary, I.J., Thriplleton, M.J., Touyz, R.M., Wardlaw, J.M., 2022.  
921 Assessment of perivascular space filtering methods using a three-dimensional  
922 computational model. *Magn Reson Imaging* 93, 33–51.  
923 <https://doi.org/10.1016/j.mri.2022.07.016>

924 Bernal, J., Xu, W., Valdés-Hernández, M. d. C., Escudero, J., Jochems, A.C.C.,  
925 Clancy, U., Doubal, F.N., Stringer, M.S., Thriplleton, M.J., Touyz, R.M., Wardlaw,  
926 J.M., 2021b. Selective Motion Artefact Reduction via Radiomics and k-space  
927 Reconstruction for Improving Perivascular Space Quantification in Brain Magnetic  
928 Resonance Imaging, *Lecture Notes in Computer Science* (including subseries

929 Lecture Notes in Artificial Intelligence and Lecture Notes in Bioinformatics).  
930 Springer International Publishing. [https://doi.org/10.1007/978-3-030-80432-9\\_12](https://doi.org/10.1007/978-3-030-80432-9_12)

931 Besteher, B., Machnik, M., Troll, M., Toepffer, A., Zerekidze, A., Rocktäschel, T.,  
932 Heller, C., Kikinis, Z., Brodoehl, S., Finke, K., Reuken, P.A., Opel, N., Stallmach,  
933 A., Gaser, C., Walter, M., 2022. Larger gray matter volumes in neuropsychiatric  
934 long-COVID syndrome. Psychiatry Res 317.  
935 <https://doi.org/10.1016/j.psychres.2022.114836>

936 Billot, B., Greve, D.N., Puonti, O., Thielscher, A., Van Leemput, K., Fischl, B., Dalca,  
937 A. V., Iglesias, J.E., 2023a. SynthSeg: Segmentation of brain MRI scans of any  
938 contrast and resolution without retraining. Med Image Anal 86.  
939 <https://doi.org/10.1016/j.media.2023.102789>

940 Billot, B., Magdamo, C., Cheng, Y., Arnold ID, S.E., Das, S.I., Eugenio Iglesias, J.,  
941 2023b. Robust machine learning segmentation for large-scale analysis of  
942 heterogeneous clinical brain MRI datasets. PNAS 120, e2216399120.  
943 <https://doi.org/https://doi.org/10.1073/pnas.2216399120>

944 Boespflug, E.L., Schwartz, D.L., Lahna, D., Pollock, J., Iliff, J.J., Kaye, J.A., Rooney,  
945 W., Silbert, L.C., 2018. MR imaging-based multimodal autoidentification of  
946 perivascular spaces (mMAPS): Automated morphologic segmentation of  
947 enlarged perivascular spaces at clinical field strength. Radiology 286, 632–642.  
948 <https://doi.org/10.1148/radiol.2017170205>

949 Boutinaud, P., Tsuchida, A., Laurent, A., Adonias, F., Hanifehliou, Z., Nozais, V.,  
950 Verrecchia, V., Lampe, L., Zhang, J., Zhu, Y.C., Tzourio, C., Mazoyer, B., Joliot,  
951 M., 2021a. 3D Segmentation of Perivascular Spaces on T1-Weighted 3 Tesla MR

952 Images With a Convolutional Autoencoder and a U-Shaped Neural Network. *Front*  
953 *Neuroinform* 15. <https://doi.org/10.3389/fninf.2021.641600>

954 Boutinaud, P., Tsuchida, A., Laurent, A., Adonias, F., Hanifehlou, Z., Nozais, V.,  
955 Verrecchia, V., Lampe, L., Zhang, J., Zhu, Y.C., Tzourio, C., Mazoyer, B., Joliot,  
956 M., 2021b. 3D Segmentation of Perivascular Spaces on T1-Weighted 3 Tesla MR  
957 Images With a Convolutional Autoencoder and a U-Shaped Neural Network. *Front*  
958 *Neuroinform* 15, 1–21. <https://doi.org/10.3389/fninf.2021.641600>

959 Bown, C.W., Carare, R.O., Schrag, M.S., Jefferson, A.L., 2022. Physiology and  
960 Clinical Relevance of Enlarged Perivascular Spaces in the Aging Brain. *Neurology*  
961 98, 107–117. <https://doi.org/10.1212/WNL.0000000000013077>

962 Braun, M., Iliff, J.J., 2020. The impact of neurovascular, blood-brain barrier, and  
963 glymphatic dysfunction in neurodegenerative and metabolic diseases, 1st ed,  
964 International Review of Neurobiology. Elsevier Inc.  
965 <https://doi.org/10.1016/bs.irn.2020.02.006>

966 Cai, D., Pan, M., Liu, C., He, W., Ge, X., Lin, J., Li, R., Liu, M., Xia, J., 2024. Deep-  
967 learning-based segmentation of perivascular spaces on T2-Weighted 3T  
968 magnetic resonance images. *Front Aging Neurosci* 16.  
969 <https://doi.org/10.3389/fnagi.2024.1457405>

970 Chai, Y., Zhang, H., Robles, C., Kim, A.S., Janhanshad, N., Thompson, P.M., van der  
971 Werf, Y., van Heese, E.M., Kim, J., Joo, E.Y., Aksman, L., Kang, K.-W., Shin, J.-  
972 W., Trang, A., Ha, J., Lee, E., Moon, Y., Kim, H., 2025. Precise perivascular space  
973 segmentation on magnetic resonance imaging from Human Connectome Project-  
974 Aging. *medRxiv* 1–11. <https://doi.org/10.1101/2025.03.19.25324269>

975 Chalcroft, L., Pappas, I., Price, C.J., Ashburner, J., 2025. Synthetic Data for Robust  
976 Stroke Segmentation. *Machine Learning for Biomedical Imaging* 3, 317–347.

977 Duarte Coello, R., Valdés Hernández, M. del C., Zwanenburg, J.J.M., van der Velden,  
978 M., Kuijf, H.J., De Luca, A., Moyano, J.B., Ballerini, L., Chappell, F.M., Brown, R.,  
979 Jan Biessels, G., Wardlaw, J.M., 2024. Detectability and accuracy of  
980 computational measurements of in-silico and physical representations of enlarged  
981 perivascular spaces from magnetic resonance images. *J Neurosci Methods* 403.  
982 <https://doi.org/10.1016/j.jneumeth.2023.110039>

983 Dubost, F., Adams, H., Bortsova, G., Ikram, M.A., Niessen, W., Vernooij, M., de  
984 Bruijne, M., 2019a. 3D regression neural network for the quantification of enlarged  
985 perivascular spaces in brain MRI. *Med Image Anal* 51, 89–100.  
986 <https://doi.org/10.1016/j.media.2018.10.008>

987 Dubost, F., Yilmaz, P., Adams, H., Bortsova, G., Ikram, M.A., Niessen, W., Vernooij,  
988 M., de Bruijne, M., 2019b. Enlarged perivascular spaces in brain MRI: Automated  
989 quantification in four regions. *Neuroimage* 185, 534–544.  
990 <https://doi.org/10.1016/j.neuroimage.2018.10.026>

991 Duering, M., Biessels, G.J., Brodtmann, A., Chen, C., Cordonnier, C., de Leeuw, F.-  
992 E., Debette, S., Frayne, R., Jouvent, E., Rost, N.S., ter Telgte, A., Al-Shahi  
993 Salman, R., Backes, W.H., Bae, H.-J., Brown, R., Chabriat, H., De Luca, A.,  
994 DeCarli, C., Dewenter, A., Doubal, F.N., Ewers, M., Field, T.S., Ganesh, A.,  
995 Greenberg, S., Helmer, K.G., Hilal, S., Jochems, A.C.C., Jokinen, H., Kuijf, H.,  
996 Lam, B.Y.K., Lebenberg, J., MacIntosh, B.J., Maillard, P., Mok, V.C.T., Pantoni,  
997 L., Rudilosso, S., Satizabal, C.L., Schirmer, M.D., Schmidt, R., Smith, C., Staals,  
998 J., Thriplleton, M.J., van Veluw, S.J., Vemuri, P., Wang, Y., Werring, D., Zedde,

999 M., Akinyemi, R.O., Del Brutto, O.H., Markus, H.S., Zhu, Y.-C., Smith, E.E.,  
1000 Dichgans, M., Wardlaw, J.M., 2023. Neuroimaging standards for research into  
1001 small vessel disease—advances since 2013. *Lancet Neurol* 22, 602–618.  
1002 [https://doi.org/10.1016/S1474-4422\(23\)00131-X](https://doi.org/10.1016/S1474-4422(23)00131-X)

1003 Francis, F., Ballerini, L., Wardlaw, J.M., 2019. Perivascular spaces and their  
1004 associations with risk factors, clinical disorders and neuroimaging features: A  
1005 systematic review and meta-analysis. *International Journal of Stroke* 14, 359–  
1006 371. <https://doi.org/10.1177/1747493019830321>

1007 Frangi, A.F., Niessen, W.J., Vincken, K.L., Viergever, M.A., 1998. Multiscale Vessel  
1008 Enhancement Filtering, in: *Lecture Notes in Computer Science*. pp. 130–137.

1009 González-Castro, V., Hernández, M.D.C.V., Armitage, P.A., Wardlaw, J.M., 2016.  
1010 Texture-based Classification for the Automatic Rating of the Perivascular Spaces  
1011 in Brain MRI, in: *Procedia Computer Science*. Elsevier B.V., pp. 9–14.  
1012 <https://doi.org/10.1016/j.procs.2016.07.003>

1013 Gouveia-Freitas, K., Bastos-Leite, A.J., 2021. Perivascular spaces and brain waste  
1014 clearance systems: relevance for neurodegenerative and cerebrovascular  
1015 pathology. *Neuroradiology*. <https://doi.org/10.1007/s00234-021-02718-7>

1016 Gudbjartsson, H., Patz, S., 1995. The Rician Distribution of Noisy MRI Data. *Magn  
1017 Reson Med* 34, 910–914.

1018 Hablitz, L.M., Nedergaard, M., 2021. The glymphatic system: A novel component of  
1019 fundamental neurobiology. *Journal of Neuroscience* 41, 7698–7711.  
1020 <https://doi.org/10.1523/JNEUROSCI.0619-21.2021>

1021 Hirschler, L., Runderkamp, B.A., Decker, A., van Harten, T.W., Scheyhing, P., Ehses,  
1022 P., Petitclerc, L., Layer, J., Pracht, E., Coolen, B.F., van der Zwaag, W., Stöcker,  
1023 T., Vollmuth, P., Paech, D., Effland, A., van Walderveen, M.A.A., Radbruch, A.,  
1024 van Buchem, M.A., Petzold, G.C., van Veluw, S.J., Caan, M.W.A., Deike, K., van  
1025 Osch, M.J.P., 2025. Region-specific drivers of CSF mobility measured with MRI  
1026 in humans. *Nat Neurosci.* <https://doi.org/10.1038/s41593-025-02073-3>

1027 Hoffmann, M., Hoopes, A., Greve, D.N., Fischl, B., Dalca, A. V., 2024. Anatomy-aware  
1028 and acquisition-agnostic joint registration with SynthMorph. *Imaging*  
1029 *Neuroscience* 2, 1–33. [https://doi.org/10.1162/imag\\_a\\_00197](https://doi.org/10.1162/imag_a_00197)

1030 Hoopes, A., Mora, J.S., Dalca, A. V., Fischl, B., Hoffmann, M., 2022. SynthStrip: skull-  
1031 stripping for any brain image. *Neuroimage* 260.  
1032 <https://doi.org/10.1016/j.neuroimage.2022.119474>

1033 Hou, Y., Park, S.H., Wang, Q., Zhang, J., Zong, X., Lin, W., Shen, D., 2017.  
1034 Enhancement of Perivascular Spaces in 7 T MR Image using Haar Transform of  
1035 Non-local Cubes and Block-matching Filtering. *Sci Rep* 7.  
1036 <https://doi.org/10.1038/s41598-017-09336-5>

1037 Iglesias, J.E., Billot, B., Balbastre, Y., Magdamo, C., Arnold, S.E., Das, S., Edlow, B.L.,  
1038 Alexander, D.C., Golland, P., Fischl, B., 2023. SynthSR: A public AI tool to turn  
1039 heterogeneous clinical brain scans into high-resolution T1-weighted images for  
1040 3D morphometry. *Sci Adv* eadd3607.

1041 Iliff, J.J., Chen, M.J., Plog, B.A., Zeppenfeld, D.M., Soltero, M., Yang, L., Singh, I.,  
1042 Deane, R., Nedergaard, M., 2014. Impairment of glymphatic pathway function  
1043 promotes tau pathology after traumatic brain injury. *Journal of Neuroscience* 34,  
1044 16180–16193. <https://doi.org/10.1523/JNEUROSCI.3020-14.2014>

1045 Iliff, J.J., Wang, M., Liao, Y., Plogg, B.A., Peng, W., Gundersen, G.A., Benveniste, H.,  
1046 Vates, G.E., Deane, R., Goldman, S.A., Nagelhus, E.A., Nedergaard, M., 2012.  
1047 A paravascular pathway facilitates CSF flow through the brain parenchyma and  
1048 the clearance of interstitial solutes, including amyloid  $\beta$ . *Sci Transl Med* 4, 1–12.  
1049 <https://doi.org/10.1126/scitranslmed.3003748>

1050 Ineichen, B. V., Okar, S. V., Proulx, S.T., Engelhardt, B., Lassmann, H., Reich, D.S.,  
1051 2022. Perivascular spaces and their role in neuroinflammation. *Neuron* 110,  
1052 3566–3581. <https://doi.org/10.1016/j.neuron.2022.10.024>

1053 Isensee, F., Jaeger, P.F., Kohl, S.A.A., Petersen, J., Maier-Hein, K.H., 2021. nnU-Net:  
1054 a self-configuring method for deep learning-based biomedical image  
1055 segmentation. *Nat Methods* 18, 203–211. [https://doi.org/10.1038/s41592-020-01008-z](https://doi.org/10.1038/s41592-020-<br/>1056 01008-z)

1057 Kern, K.C., Nasrallah, I.M., Bryan, R.N., Reboussin, D.M., Wright, C.B., 2023.  
1058 Intensive systolic blood pressure treatment remodels brain perivascular spaces:  
1059 A secondary analysis of the Systolic Pressure Intervention Trial (SPRINT).  
1060 *Neuroimage Clin* 40, 103513. <https://doi.org/10.1016/j.nicl.2023.103513>

1061 Kim, H.G., Shin, N.-Y., Nam, Y., Yun, E., Yoon, U., Lee, H.S., Ahn, K.J., 2023. MRI-  
1062 visible Dilated Perivascular Space in the Brain by Age: The Human Connectome  
1063 Project. *Radiology* 306, 1–9. <https://doi.org/10.1148/radiol.213254>

1064 Laso, P., Cerri, S., Sorby-Adams, A., Guo, J., Mateen, F., Goebl, P., Wu, J., Liu, P.,  
1065 Li, H., Young, S.I., Billot, B., Puonti, O., Sze, G., Payabavash, S., DeHavenon, A.,  
1066 Sheth, K.N., Rosen, M.S., Kirsch, J., Strisciuglio, N., Wolterink, J.M., Eshaghi, A.,  
1067 Barkhof, F., Kimberly, W.T., Iglesias, J.E., 2024. Quantifying white matter  
1068 hyperintensity and brain volumes in heterogeneous clinical and low-field portable

1069 MRI, in: IEEE International Symposium on Biomedical Imaging (ISBI). IEEE,  
1070 Athens, Greece, pp. 1–5.

1071 Lian, C., Zhang, J., Liu, M., Zong, X., Hung, S.C., Lin, W., Shen, D., 2018. Multi-  
1072 channel multi-scale fully convolutional network for 3D perivascular spaces  
1073 segmentation in 7T MR images. *Med Image Anal* 46, 106–117.  
1074 <https://doi.org/10.1016/j.media.2018.02.009>

1075 Lynch, K.M., Sepehrband, F., Toga, A.W., Choupan, J., 2023. Brain perivascular  
1076 space imaging across the human lifespan. *Neuroimage* 271, 120009.  
1077 <https://doi.org/10.1016/j.neuroimage.2023.120009>

1078 Maier-Hein, L., Reinke, A., Godau, P., Tizabi, M.D., Buettner, F., Christodoulou, E.,  
1079 Glocker, B., Isensee, F., Kleesiek, J., Kozubek, M., Reyes, M., Riegler, M.A.,  
1080 Wiesenfarth, M., Kavur, A.E., Sudre, C.H., Baumgartner, M., Eisenmann, M.,  
1081 Heckmann-Nötzel, D., Rädsch, T., Acion, L., Antonelli, M., Arbel, T., Bakas, S.,  
1082 Benis, A., Blaschko, M.B., Cardoso, M.J., Cheplygina, V., Cimini, B.A., Collins,  
1083 G.S., Farahani, K., Ferrer, L., Galdran, A., van Ginneken, B., Haase, R.,  
1084 Hashimoto, D.A., Hoffman, M.M., Huisman, M., Jannin, P., Kahn, C.E.,  
1085 Kainmueller, D., Kainz, B., Karargyris, A., Karthikesalingam, A., Kofler, F., Kopp-  
1086 Schneider, A., Kreshuk, A., Kurc, T., Landman, B.A., Litjens, G., Madani, A.,  
1087 Maier-Hein, K., Martel, A.L., Mattson, P., Meijering, E., Menze, B., Moons, K.G.M.,  
1088 Müller, H., Nichyporuk, B., Nickel, F., Petersen, J., Rajpoot, N., Rieke, N., Saez-  
1089 Rodriguez, J., Sánchez, C.I., Shetty, S., van Smeden, M., Summers, R.M., Taha,  
1090 A.A., Tiulpin, A., Tsaftaris, S.A., Van Calster, B., Varoquaux, G., Jäger, P.F.,  
1091 2024. Metrics reloaded: recommendations for image analysis validation. *Nat  
1092 Methods* 21, 195–212. <https://doi.org/10.1038/s41592-023-02151-z>

1093 Menze, I., Bernal, J., Kaya, P., Aki, Ç., Pfister, M., Geisendorfer, J., Yakupov, R.,  
1094 Coello, R.D., Valdés-Hernández, M. d. C., Heneka, M.T., Brosseron, F., Schmid,  
1095 M.C., Glanz, W., Incesoy, E.I., Butryn, M., Rostamzadeh, A., Meiberth, D., Peters,  
1096 O., Preis, L., Lammerding, D., Gref, D., Priller, J., Spruth, E.J., Altenstein, S.,  
1097 Lohse, A., Hetzer, S., Schneider, A., Fliessbach, K., Kimmich, O., Vogt, I.R.,  
1098 Wiltfang, J., Bartels, C., Schott, B.H., Hansen, N., Dechent, P., Buerger, K.,  
1099 Janowitz, D., Perneczky, R., Rauchmann, B.-S., Teipel, S., Kilimann, I., Goerss,  
1100 D., Laske, C., Munk, M.H., Sanzenbacher, C., Hinderer, P., Scheffler, K., Spottke,  
1101 A., Roy-Kluth, N., Lüsebrink, F., Neumann, K., Wardlaw, J., Jessen, F., Schreiber,  
1102 S., Düzel, E., Ziegler, G., 2024. Perivascular space enlargement accelerates in  
1103 ageing and Alzheimer's disease pathology: evidence from a three-year  
1104 longitudinal multicentre study. *Alzheimers Res Ther* 16, 242.  
1105 <https://doi.org/10.1186/s13195-024-01603-8>

1106 Merveille, O., Talbot, H., Najman, L., Passat, N., 2018. Curvilinear Structure Analysis  
1107 by Ranking the Orientation Responses of Path Operators. *IEEE Trans Pattern  
1108 Anal Mach Intell* 40, 304–317. <https://doi.org/10.1109/TPAMI.2017.2672972>

1109 Merveille, O., Talbot, H., Najman, L., Passat, N., 2014. Tubular structure filtering by  
1110 ranking orientation responses of path operators. *Lecture Notes in Computer  
1111 Science (including subseries Lecture Notes in Artificial Intelligence and Lecture  
1112 Notes in Bioinformatics)* 8690 LNCS, 203–218. [319-10605-2\\_14](https://doi.org/10.1007/978-3-<br/>1113 319-10605-2_14)

1114 Mestre, H., Tithof, J., Du, T., Song, W., Peng, W., Sweeney, A.M., Olveda, G.,  
1115 Thomas, J.H., Nedergaard, M., Kelley, D.H., 2018. Flow of cerebrospinal fluid is

1116 driven by arterial pulsations and is reduced in hypertension. *Nat Commun* 9, 4878.

1117 <https://doi.org/10.1038/s41467-018-07318-3>

1118 Milletari, F., Navab, N., Ahmadi, S.-A., 2016. V-Net: Fully Convolutional Neural  
1119 Networks for Volumetric Medical Image Segmentation, in: 2016 Fourth  
1120 International Conference on 3D Vision (3DV). pp. 565–571.

1121 Müller, P., Horndasch, L., Neumann, K., Mattern, H., Cardace, S., Arndt, P., Pfister,  
1122 M., Groscheck, T., Vielhaber, S., Meuth, S., Dunay, I., Schmeisser, A., Behme,  
1123 D., Schreiber, S., Braun-dullaes Rüdiger, 2024. CEREBRAL SMALL VESSEL  
1124 DISEASE MEDIATES THE EFFECT OF ARTERIAL STIFFNESS ON  
1125 COGNITIVE DECLINE IN PATIENTS WITH HEART FAILURE WITH  
1126 PRESERVED EJECTION FRACTION. *J Hypertens* 42(Suppl 1), e92–e93.

1127 Neumann, K., Günther, M., Düzel, E., Schreiber, S., 2022. Microvascular Impairment  
1128 in Patients With Cerebral Small Vessel Disease Assessed With Arterial Spin  
1129 Labeling Magnetic Resonance Imaging: A Pilot Study. *Front Aging Neurosci* 14.  
1130 <https://doi.org/10.3389/fnagi.2022.871612>

1131 Okar, S. V., Hu, F., Shinohara, R.T., Beck, E.S., Reich, D.S., Ineichen, B. V., 2023.  
1132 The etiology and evolution of magnetic resonance imaging-visible perivascular  
1133 spaces: Systematic review and meta-analysis. *Front Neurosci* 17, 1–13.  
1134 <https://doi.org/10.3389/fnins.2023.1038011>

1135 Park, S.H., Zong, X., Gao, Y., Lin, W., Shen, D., 2016. Segmentation of perivascular  
1136 spaces in 7 T MR image using auto-context model with orientation-normalized  
1137 features. *Neuroimage* 134, 223–235.  
1138 <https://doi.org/10.1016/j.neuroimage.2016.03.076>

1139 Pham, W., Jarema, A., Rim, D., Chen, Z., Khelif, M.S., Macefield, V.G., Henderson,  
1140 L.A., Brodtmann, A., 2024. A Comprehensive Framework for Automated  
1141 Segmentation of Perivascular Spaces in Brain MRI with the nnU-Net. ArXiv 1–49.

1142 Pham, W., Lynch, M., Spitz, G., O'Brien, T., Vivash, L., Sinclair, B., Law, M., 2022. A  
1143 critical guide to the automated quantification of perivascular spaces in magnetic  
1144 resonance imaging. *Front Neurosci* 16.  
1145 <https://doi.org/10.3389/fnins.2022.1021311>

1146 Potter, G.M., Chappell, F.M., Morris, Z., Wardlaw, J.M., 2015. Cerebral perivascular  
1147 spaces visible on magnetic resonance imaging: Development of a qualitative  
1148 rating scale and its observer reliability. *Cerebrovascular Diseases* 39, 224–231.  
1149 <https://doi.org/10.1159/000375153>

1150 Rashid, T., Liu, H., Ware, J.B., Li, K., Romero, J.R., Fadaee, E., Nasrallah, I.M., Hilal,  
1151 S., Bryan, R.N., Hughes, T.M., Davatzikos, C., Launer, L., Seshadri, S., Heckbert,  
1152 S.R., Habes, M., 2023. Deep learning based detection of enlarged perivascular  
1153 spaces on brain MRI. *Neuroimage: Reports* 3.  
1154 <https://doi.org/10.1016/j.ynirp.2023.100162>

1155 Rasmussen, M.K., Mestre, H., Nedergaard, M., 2018. The glymphatic pathway in  
1156 neurological disorders. *Lancet Neurol* 17, 1016–1024.  
1157 [https://doi.org/10.1016/S1474-4422\(18\)30318-1](https://doi.org/10.1016/S1474-4422(18)30318-1)

1158 Saito, T., Rehmsmeier, M., 2015. The precision-recall plot is more informative than the  
1159 ROC plot when evaluating binary classifiers on imbalanced datasets. *PLoS One*  
1160 10. <https://doi.org/10.1371/journal.pone.0118432>

1161 Schreiber, S., Bernal, J., Arndt, P., Schreiber, F., Müller, P., Morton, L., Braun-  
1162 Dullaeus, R.C., Valdés-Hernández, M.D.C., Duarte, R., Wardlaw, J.M., Meuth,

1163 S.G., Mietzner, G., Vielhaber, S., Dunay, I.R., Dityatev, A., Jandke, S., Mattern,  
1164 H., 2023. Brain Vascular Health in ALS Is Mediated through Motor Cortex  
1165 Microvascular Integrity. *Cells* 12. <https://doi.org/10.3390/cells12060957>

1166 Schwartz, D.L., Boespflug, E.L., Lahna, D.L., Pollock, J., Roese, N.E., Silbert, L.C.,  
1167 2019. Autoindentification of perivascular spaces in white matter using clinical field  
1168 strength T1 and FLAIR MR imaging. *Neuroimage* 202, 116126.  
1169 <https://doi.org/10.1016/j.neuroimage.2019.116126>

1170 Shaw, R., Sudre, C.H., Varsavsky, T., Ourselin, S., Cardoso, M.J., 2020. A k-Space  
1171 Model of Movement Artefacts: Application to Segmentation Augmentation and  
1172 Artefact Removal. *IEEE Trans Med Imaging* 39, 2881–2892.  
1173 <https://doi.org/10.1109/tmi.2020.2972547>

1174 Smith, E.E., Biessels, G.J., De Guio, F., de Leeuw, F.E., Duchesne, S., Düring, M.,  
1175 Frayne, R., Ikram, M.A., Jouvent, E., MacIntosh, B.J., Thriplleton, M.J., Vernooij,  
1176 M.W., Adams, H., Backes, W.H., Ballerini, L., Black, S.E., Chen, C., Corriveau,  
1177 R., DeCarli, C., Greenberg, S.M., Gurol, M.E., Ingrisch, M., Job, D., Lam, B.Y.K.,  
1178 Launer, L.J., Linn, J., McCreary, C.R., Mok, V.C.T., Pantoni, L., Pike, G.B.,  
1179 Ramirez, J., Reijmer, Y.D., Romero, J.R., Ropele, S., Rost, N.S., Sachdev, P.S.,  
1180 Scott, C.J.M., Seshadri, S., Sharma, M., Sourbron, S., Steketee, R.M.E., Swartz,  
1181 R.H., van Oostenbrugge, R., van Osch, M., van Rooden, S., Viswanathan, A.,  
1182 Werring, D., Dichgans, M., Wardlaw, J.M., 2019. Harmonizing brain magnetic  
1183 resonance imaging methods for vascular contributions to neurodegeneration.  
1184 *Alzheimer's and Dementia: Diagnosis, Assessment and Disease Monitoring* 11,  
1185 191–204. <https://doi.org/10.1016/j.dadm.2019.01.002>

1186 Sudre, C.H., Van Wijnen, K., Dubost, F., Adams, H., Atkinson, D., Barkhof, F., Birhanu,  
1187 M.A., Bron, E.E., Camarasa, R., Chaturvedi, N., Chen, Y., Chen, Z., Chen, S.,  
1188 Dou, Q., Evans, T., Ezhov, I., Gao, H., Girones Sanguesa, M., Gispert, J.D.,  
1189 Gomez Anson, B., Hughes, A.D., Ikram, M.A., Ingala, S., Jaeger, H.R., Kofler, F.,  
1190 Kuijf, H.J., Kutnar, D., Lee, M., Li, B., Lorenzini, L., Menze, B., Molinuevo, J.L.,  
1191 Pan, Y., Puybareau, E., Rehwald, R., Su, R., Shi, P., Smith, L., Tillin, T., Tochon,  
1192 G., Urien, H., van der Velden, B.H.M., van der Velpen, I.F., Wiestler, B., Wolters,  
1193 F.J., Yilmaz, P., de Groot, M., Vernooij, M.W., de Brujne, M., 2024. Where is  
1194 VALDO? VAscular Lesions Detection and segmentatiOn challenge at MICCAI  
1195 2021. *Med Image Anal* 91. <https://doi.org/10.1016/j.media.2023.103029>

1196 Tobin, J., Fong, R., Ray, A., Schneider, J., Zaremba, W., Abbeel, P., 2017. Domain  
1197 Randomization for Transferring Deep Neural Networks from Simulation to the  
1198 Real World, in: IEEE/RSJ International Conference on Intelligent Robots and  
1199 Systems (IROS). Vancouver, pp. 23–30.  
1200 <https://doi.org/10.1109/IROS.2017.8202133>

1201 Valdes Hernandez, M. d. C., Piper, R.J., Wang, X., Deary, I.J., Wardlaw, J.M., 2013.  
1202 Towards the automatic computational assessment of enlarged perivascular  
1203 spaces on brain magnetic resonance images: A systematic review. *Journal of*  
1204 *Magnetic Resonance Imaging* 38, 774–785. <https://doi.org/10.1002/jmri.24047>

1205 Valdés Hernández, M. del C., Duarte Coello, R., Xu, W., Bernal, J., Cheng, Y.,  
1206 Ballerini, L., Wiseman, S.J., Chappell, F.M., Clancy, U., Jaime García, D., Arteaga  
1207 Reyes, C., Zhang, J.F., Liu, X., Hewins, W., Stringer, M., Doubal, F., Thrippleton,  
1208 M.J., Jochems, A., Brown, R., Wardlaw, J.M., 2024. Influence of threshold  
1209 selection and image sequence in in-vivo segmentation of enlarged perivascular

1210 spaces. J Neurosci Methods 403.

1211 <https://doi.org/10.1016/j.jneumeth.2023.110037>

1212 Vikner, T., Karalija, N., Eklund, A., Malm, J., Lundquist, A., Gallewicz, N., Dahlin, M.,  
1213 Lindenberger, U., Riklund, K., Bäckman, L., Nyberg, L., Wåhlin, A., 2022. 5-Year  
1214 Associations among Cerebral Arterial Pulsatility, Perivascular Space Dilation, and  
1215 White Matter Lesions. Ann Neurol 92, 871–881.

1216 <https://doi.org/10.1002/ana.26475>

1217 Wardlaw, J.M., Benveniste, H., Nedergaard, M., Zlokovic, B. V., Mestre, H., Lee, H.,  
1218 Doubal, F.N., Brown, R., Ramirez, J., MacIntosh, B.J., Tannenbaum, A., Ballerini,  
1219 L., Rungta, R.L., Boido, D., Sweeney, M., Montagne, A., Charpak, S., Joutel, A.,  
1220 Smith, K.J., Black, S.E., 2020. Perivascular spaces in the brain: anatomy,  
1221 physiology and pathology. Nat Rev Neurol 16, 137–153.

1222 <https://doi.org/10.1038/s41582-020-0312-z>

1223 Wardlaw, J.M., Doubal, F., Armitage, P., Chappell, F., Carpenter, T., Muñoz Maniega,  
1224 S., Farrall, A., Sudlow, C., Dennis, M., Dhillon, B., 2009. Lacunar stroke is  
1225 associated with diffuse Blood-Brain barrier dysfunction. Ann Neurol 65, 194–202.

1226 <https://doi.org/10.1002/ana.21549>

1227 Waymont, J.M.J., Valdés Hernández, M. del C., Bernal, J., Duarte Coello, R., Brown,  
1228 R., Chappell, F.M., Ballerini, L., Wardlaw, J.M., 2024. Systematic review and  
1229 meta-analysis of automated methods for quantifying enlarged perivascular  
1230 spaces in the brain. Neuroimage.

1231 <https://doi.org/10.1016/j.neuroimage.2024.120685>

1232 Wiltgen, T., McGinnis, J., Schlaeger, S., Kofler, F., Voon, C.C., Berthele, A., Bischl,  
1233 D., Grundl, L., Will, N., Metz, M., Schinz, D., Sepp, D., Prucker, P., Schmitz-Koep,

1234 B., Zimmer, C., Menze, B., Rueckert, D., Hemmer, B., Kirschke, J., Mühlau, M.,  
1235 Wiestler, B., 2024. LST-AI: A deep learning ensemble for accurate MS lesion  
1236 segmentation. *Neuroimage Clin* 42. <https://doi.org/10.1016/j.nicl.2024.103611>

1237 Yamamoto, E.A., Bagley, J.H., Geltzeiler, M., Sanusi, O.R., Dogan, A., Liu, J.J.,  
1238 Piantino, J., 2024. The perivascular space is a conduit for cerebrospinal fluid flow  
1239 in humans: A proof-of-principle report. *Proc Natl Acad Sci U S A* 121.  
1240 <https://doi.org/10.1073/pnas.2407246121>

1241 Zhang, J., Gao, Y., Park, S.H., Zong, X., Lin, W., Shen, D., 2017. Structured Learning  
1242 for 3-D Perivascular Space Segmentation Using Vascular Features. *IEEE Trans  
1243 Biomed Eng* 64, 2803–2812. <https://doi.org/10.1109/TBME.2016.2638918>

1244

1245